



HAL
open science

Molecular, isotopic and radiocarbon evidence for broomcorn millet cropping in Northeast France since the Bronze Age

Blandine Courel, Philippe Schaeffer, Pierre Adam, Estelle Motsch, Quentin Ebert, Emile Moser, Clément Féliu, Stefano M. Bernasconi, Irka Hajdas, Ertlen Damien, et al.

► **To cite this version:**

Blandine Courel, Philippe Schaeffer, Pierre Adam, Estelle Motsch, Quentin Ebert, et al.. Molecular, isotopic and radiocarbon evidence for broomcorn millet cropping in Northeast France since the Bronze Age. *Organic Geochemistry*, 2017, 110, pp.13-24. 10.1016/j.orggeochem.2017.03.002 . hal-01800645

HAL Id: hal-01800645

<https://inrap.hal.science/hal-01800645>

Submitted on 21 Aug 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 Molecular, isotopic and radiocarbon evidence for broomcorn millet
2 cropping in Northeast France since the Bronze Age

3 Blandine Courel^a, Philippe Schaeffer^{a*}, Pierre Adam^a, Estelle Motsch^a, Quentin Ebert^a, Emile
4 Moser^a, Clément Féliu^{b,c}, Stefano M. Bernasconi^d, Irka Hajdas^e, Damien Ertlen^f, Dominique
5 Schwartz^f

6 ^a *Université de Strasbourg, CNRS, CHIMIE, UMR 7177, F-67000 Strasbourg, France*

7 ^b *Inrap Grand-Est Sud, 67100 Strasbourg, France*

8 ^c *Université de Strasbourg, Université de Haute Alsace, CNRS, ArchIMèdE UMR 7044, F-
9 67000 Strasbourg, France*

10 ^d *Geologisches Institut, ETH Zürich, Zürich, Switzerland*

11 ^e *Laboratory of Ion Beam Physics, ETH Zürich, Zürich, Switzerland*

12 ^f *Université de Strasbourg, CNRS, Laboratoire Image, Ville, Environnement UMR 7362, F-
13 67000 Strasbourg, France*

14

15

16

17 *Corresponding author. Tel: +33 (0)3 68 85 28 05.

18 *E-mail address: p.schaeff@unistra.fr (Philippe Schaeffer).*

19 ABSTRACT

20 Molecular and isotopic investigation of lipids from soils filling several structures from an
21 archaeological site located at Obernai (Alsace, NE France) has revealed the presence of miliacin,
22 a triterpenoid marker from *Panicum miliaceum* (broomcorn millet), indicating that this cereal
23 was cultivated at the site. The concentration profiles of miliacin within silos and its detection
24 in other archaeological structures (e.g. Gaulish pit) suggest that miliacin did not originate from
25 cereals stored in the silos but rather came from remains of millet from cultivated soils which
26 filled the silos after they were abandoned. Furthermore, the ^{14}C age of miliacin isolated from a
27 silo of the Second Iron Age was shown to be considerably older (Bronze Age) than the structure
28 itself, revealing that the soil filling the silo therefore archived the molecular signature from past
29 millet cropping, predating the digging of the silo. Thus, radiocarbon dating of the isolated
30 miliacin allowed the timing of millet cropping to be determined, showing that it was established
31 during the Bronze Age and the Roman Gaul period at Obernai. This is the first evidence of
32 millet cultivation in Alsace dating back to the Bronze Age, bringing new perspectives on
33 agricultural and past dietary practices in Eastern France. The combination of molecular studies
34 and radiocarbon dating of individual lipids highlights the potential of hollow structures like
35 silos and pits to act as “pedological traps”, recording information on past vegetation cover or
36 agricultural practices from the surface horizons of surrounding soils that filled these structures
37 after abandonment.

38

39 Keywords (3-7 words): *Panicum miliaceum*; miliacin; compound-specific radiocarbon
40 analysis; archaeometry; soil lipids; pedological traps.

41 **1. Introduction**

42 When chemical analytical techniques were initially introduced to archaeological research
43 during the middle of the 20th century in a discipline referred to as archaeometry, the analysis
44 mainly involved characterization of organic residues and materials related to artifacts such as
45 pottery and jars (Condamin et al., 1976; Evershed et al., 1991; Charters et al., 1995; Charrié-
46 Duhaut et al., 2007), hafting material on flints (Boëda et al., 1996) or mummies (Connan, 1999;
47 Buckley and Evershed, 2001). It is only more recently that lipid biomarkers in organic matter
48 (OM) preserved in soils and sediments have been considered as archaeological archives (e.g.
49 Bull et al., 2001; Egli et al., 2013; Motuzaite-Matuzeviciute et al., 2016). These lipids, which
50 possess relative stability and resistance towards biodegradation (Eglinton and Logan, 1991;
51 Lorenz et al., 2007), have been shown to have a potential as chemotaxonomic markers, allowing
52 a link to be established between the preserved molecules ("chemical fossils") and their source
53 organisms. Thus, lipid biomarkers, preserved within palaeosoils and sediments, may represent
54 valuable tools for reconstructing palaeoenvironments or studying the evolution of landscapes
55 through time (Trendel et al., 2010; Lavrieux et al., 2012; Ertlen et al., 2015), or investigating
56 ancient dietary and agricultural practices (Bull et al., 1999; Jacob et al., 2008a; Sistiaga et al.,
57 2014; Motuzaite-Matuzeviciute et al., 2016) or other human activities such as hemp retting
58 (Lavrieux et al., 2013).

59 In studies dealing with soils, the lipid compartment of soil OM (SOM) is a result of the
60 combination of old and refractory compounds mixed with more recent lipid inputs originating
61 from the vegetation cover of the topsoil and introduced into deeper horizons by processes such
62 as bioturbation, leaching or root penetration (Kögel-Knabner, 2002; Schmidt et al., 2011).
63 Furthermore, the role of microorganisms living within the soil ecosystem has to be considered
64 since they consume and produce part of the soil lipid components (Kögel-Knabner, 2002). The
65 lipid content of SOM also depends strongly on the degree of preservation, which is related to

66 different factors such as climate, biological activity, pH, level of oxygenation, depth,
67 mineralogy, or biochemical nature of the soil (Schmidt et al., 2011). As a result of the intense
68 reshuffling of SOM and of the differential inherent persistence of the various lipid constituents
69 of soils, interpretation of the lipid signatures from soils in terms of archaeological significance
70 and their relationship with a chronological sequence is difficult to establish (Schwartz, 2012).
71 In this respect, radiocarbon dating of SOM, lipid fractions and, in more rare instances,
72 individual compounds from soils represents a powerful tool, potentially allowing, in the case
73 of lipids, specific molecular signatures to be related to a chronological/historical context (e.g.,
74 Rethemeyer et al., 2005; Mendez-Milan et al., 2014). The tremendous advances in ¹⁴C
75 instrumentation during the last decade enable samples containing only minute amounts of
76 carbon to be dated (e.g., Ingalls and Pearson, 2005; Birkholz et al., 2013; Gierga et al., 2016).
77 In particular, the accelerator mass spectrometry (AMS) facility MICADAS (Mini CARbon
78 DAting System), designed and developed at the ETH of Zurich, is equipped with a gas handling
79 system and a gas ion source, allowing direct radiocarbon measurement of CO₂ obtained via
80 oxidation of very small samples, thereby giving access to radiocarbon dating of single
81 compounds at the microgram scale (Ruff et al., 2007, 2010).

82 In the present study, we have investigated the content of lipid extracts (LEs) from soil material
83 filling archaeological structures (silos, pit and latrines) near the city of Obernai (Alsace, NE
84 France) using gas chromatography-MS (GC-MS) and GC-combustion- isotope ratio MS (GC-
85 *c*-IRMS). The investigation, which was originally aimed at determining the nature of the
86 cereals/plants that could have been stored in the silos and potentially cultivated at the site, has
87 revealed the presence of miliacin, a triterpenoid marker from *Panicum miliaceum* (broomcorn
88 millet). Its occurrence and significance in terms of preservation and timing of millet cultivation
89 is discussed, notably in the light of the stable carbon isotopic composition of lipids and
90 radiocarbon dating of isolated miliacin.

91 **2. Material and methods**

92 *2.1. Samples and archaeological context*

93 The archaeological site of Obernai (Alsace, NE France), spread over more than 7.5 hectares,
94 was excavated in 2013 by the Institut National de Recherche Archéologique Préventive
95 (INRAP) and has yielded numerous pieces of evidence for successive periods of human
96 occupation dating back from the Neolithic period up to the beginning of the Middle Ages (Féliu
97 et al., unpublished results). Among the structures unearthed, some corresponded to protohistoric
98 silos deeply buried in the loess substrate (ca. 1-2.2 m high by ca. 1.5-2 m wide; Fig. 1 and Fig.
99 S1 in Supplementary data). The silos, which were used for cereal storage, indicate that local
100 agricultural activity occurred at the site. Some of the silos, dated from the Late Bronze Age
101 (950-800 BC) and Second Iron Age (La Tène B, 400-260 BC), based on the typology of the
102 ceramics in the soils filling the silos, were selected for biomarker analysis (Table 1). In addition,
103 soil samples from one Neolithic silo (4600-4300 BC), one silo dated from La Tène D (150-30
104 BC), as well as from a Gaulish pit encircling a farm and Roman Gaul latrines (end of the 2nd
105 century AD – beginning of the 3rd century AD) were collected (Table 1). In the case of the
106 silos, soil samples were collected at different depth levels within the apparent archaeological
107 layers (ALs; Fig. 1 and Fig. S1 in Supplementary data). For comparison, the loess soils within
108 which the silos were dug - named "loess substrate" (LS)- were sampled.

109 *2.2. Reference material from P. miliaceum*

110 Lipids from reference plant samples from *P. miliaceum* (broomcorn millet) provided by J. Jacob
111 (Institut des Sciences de la Terre d'Orléans) were investigated. They consisted of seeds, stems,
112 ears, leaves and roots obtained from millet cultivated in the frame of a detailed study of millet
113 (Bossard et al., 2013).

114 2.3. *Cautions taken during the sample treatment*

115 In order to avoid contaminations (e.g., phthalates), all commercial solvents were redistilled and
116 stored in glass bottles carefully cleaned and closed with an aluminium foil. Prior to use, all
117 glasswares were washed using water, acetone and finally with redistilled dichloromethane. All
118 equipments/supplies (pipettes, sand, silica, celilte) were washed with dichloromethane in a
119 Soxhlet. Once cleaned, the silica gel was activated at 120 °C. Each samples was kept in a freezer
120 at -20 °C.

121 2.4. *Lipid extraction and fractionation (general procedure)*

122 Lipids from archaeological soils (ca. 500 g) and from plant reference material were extracted
123 via sonication using CH₂Cl₂/CH₃OH (1:1, v/v) followed by filtration of the supernatant through
124 celite and solvent removal under reduced pressure. In the case of archaeological soils, the yield
125 of lipid extract (LE) was low ($\leq 0.04\%$ of soil). Derivatization and fractionation of an aliquot
126 of the extract was carried out after Schnell et al. (2014). Briefly, following acetylation of the
127 alcohol functionalities using an excess of Ac₂O and 20 μ l of *N*-methylimidazole (catalyst) and
128 methylation of the carboxylic acids using CH₂N₂ in Et₂O, the derivatized crude extract was
129 fractionated on a silica gel column into an apolar fraction eluted with CH₂Cl₂/EtOAc (8:2, v/v)
130 and a more polar fraction eluted with CH₂Cl₂/CH₃OH (1:1, v/v), the latter not being investigated
131 further. After solvent extraction and prior to derivatization and fractionation, the propyl ester
132 of stearic acid was added as internal standard (IS) for quantitative analysis of miliacin.

133 2.5. *Lipid fractionation and derivatization for stable carbon isotope measurements*

134 2.5.1. *Fractionation of the LE*

135 An aliquot of the underivatized lipid extract adsorbed onto silica gel was fractionated using
136 activated silica gel column chromatography, eluting successively with cyclohexane/CH₂Cl₂

137 (9:1, v/v; 1 dead volume, V_0) to yield the hydrocarbon fraction, followed by CH_2Cl_2 ($2 \times V_0$)
138 to provide mainly miliacin, then $\text{CH}_2\text{Cl}_2/\text{EtOAc}$ (9:1, v/v; $2 \times V_0$) for the *n*-alcohols and sterols.
139 Finally, the most polar components, comprising carboxylic acids, were recovered with
140 $\text{CH}_2\text{Cl}_2/\text{CH}_3\text{OH}$ (1:1, v/v; $2 \times V_0$).

141 2.5.2. Derivatization

142 The alcohol fraction, dissolved in CH_2Cl_2 , was derivatized using pyridine (40 μl) and *bis*-(*N,O*-
143 trimethylsilyl)trifluoroacetamide (BSTFA; 70 °C, 2 h), the $\delta^{13}\text{C}$ composition of the
144 trimethylsilyl (TMS) group having been determined previously. The solvent and excess reagent
145 were removed under a stream of N_2 .

146 The polar fraction was reacted with $\text{BF}_3/\text{CH}_3\text{OH}$ (the $\delta^{13}\text{C}$ value of CH_3OH being known) at
147 60 °C for 2 h. The crude mixture was then transferred to a separatory funnel and the organic
148 layer recovered after addition of distilled water and CH_2Cl_2 . After removal of CH_2Cl_2 under
149 reduced pressure, the derivatized extract was filtered through a small silica gel column/pipette
150 using CH_2Cl_2 , yielding the fatty acid (FA) methyl ester fraction.

151 The known isotopic composition of the added TMS group ($\delta^{13}\text{C}$, -20.8‰) and CH_3OH group
152 ($\delta^{13}\text{C}$, -48.5‰) allowed the $\delta^{13}\text{C}$ value of the derivatized lipids to be corrected using the
153 following equation (Rieley, 1994):

$$154 \quad \delta_c = (n_{cd}\delta_{cd} - n_d\delta_d) / n_c$$

155 with *n* the number of carbon and δ the carbon isotopic ratio for derivatized compounds (*cd*), the
156 derivative group added (*d*) and the compound prior to derivatization (*c*).

157 2.6. Decarbonation of soil samples

158 For radiocarbon dating and $\delta^{13}\text{C}$ determination of SOM, an aliquot of the soil sample was
159 decarbonated using 6 N HCl at room temperature (15 min). After centrifugation, the supernatant
160 was removed by pipetting and the residue rinsed with distilled water. The procedure was
161 repeated until neutral and the residue oven-dried at 50 °C.

162 2.7. Isolation of miliacin

163 The LE from selected samples (Table 1) was fractionated on a silica gel column, eluting
164 successively with cyclohexane/ CH_2Cl_2 (8:2), CH_2Cl_2 and $\text{CH}_2\text{Cl}_2/\text{CH}_3\text{OH}$ (1:1), the second-
165 eluted fraction being enriched in miliacin, as shown from GC-MS analysis (Fig. S2 in
166 Supplementary data). This fraction was further purified and manually collected using high
167 performance liquid chromatography (HPLC) with a Waters model 590 HPLC pump (mobile
168 phase: $\text{CH}_2\text{Cl}_2/\text{CH}_3\text{OH}$, 2:8, v/v; flow rate, 1 ml/min), connected to an Agilent ZORBAX SB-
169 C18 column (25 cm x 4.6 mm i.d., 5 μm , Agilent) and associated with a differential
170 refractometer detector R401 (Waters Associates). After a control of the purity of the miliacin
171 sample using GC flame ionization detection (GC-FID) , a final clean up step using silica gel
172 thin layer chromatography (TLC, 0.5 mm SiO_2) was sometimes required (samples 1330.AL2,
173 6, 10) and carried out using cyclohexane/ Me_2CO (98:2) as developer, miliacin having a
174 retention factor between 0.76 and 0.83. The final purity was checked using GC-FID and GC-
175 MS (Fig. S2, Supplementary data).

176 2.8. GC-MS

177 GC-MS was carried out with a Thermo Trace gas chromatograph (Thermo Scientific) equipped
178 with Tri Plus autosampler, a programmed temperature vaporizing (PTV) injector and a HP5-
179 MS column (30 m x 0.25 mm i.d. x 0.25 μm film thickness) using He as carrier gas (constant
180 1.1 ml/min). The temperature program used a gradient from 70 °C to 200 °C at 10 °C/min, then
181 from 200 °C to 300 °C at 3 °C/min (conditions A) or at 4°C/min (conditions B) followed by

182 isothermal at 300 °C for 40 min. The GC equipment was coupled to a Thermo TSQ Quantum
183 mass spectrometer working in the electron ionization (EI) mode at 70 eV and scanning m/z 50
184 to 700. The data were investigated using the Xcalibur Software and mass spectra were
185 compared with the NIST library and literature data (for MS data, see Table S1 in Supplementary
186 data).

187 2.9. GC-FID

188 GC-FID was carried out with a HEWLETT-PACKARD HP 6890 gas chromatograph equipped
189 with an on-column injector, FID and a HP5-MS column (30 m x 0.32 mm i.d. x 0.25 μm film
190 thickness). The temperature program was: 70 °C – 300 °C (held 40 min) at 10 °C/min), with H₂
191 as carrier gas (constant 2.5 ml/min). The temperature of the detector was 310 °C.

192 2.10. Elementar analysis-combustion-isotope ratio mass spectrometry (EA-c-IRMS)

193 Bulk carbon isotope composition ($\delta^{13}\text{C}$) of SOM was determined using an Elementar Vario
194 Micro Cube equipped with thermal conductivity detection and coupled to an Isoprime visION
195 stable isotope ratio mass spectrometer. Homogenized decarbonated soil samples were weighted
196 into Sn capsules using a microbalance ($2 \times 10^{-2}\text{g}$; CPA26P Sartorius). The capsules were
197 individually introduced into a combustion furnace (950 °C) with an excess of O₂. CuO was used
198 as the oxidation catalyst and He was the carrier gas. Water was removed with a P₂O₅ trap and
199 CO₂ was separated with a purge and trap desorption column. Ion currents (m/z 44, 45, 46) were
200 measured continuously for CO₂ using triple Faraday cups connected to high gain amplifiers.
201 Reference pulse peaks of laboratory CO₂ gas were calibrated against the international standards
202 IAEA C3 ($\delta^{13}\text{C} = -24.91$ (0.49) ‰) and IAEA C4 ($\delta^{13}\text{C} = -23.96$ (0.62) ‰) for CO₂ (IAEA,
203 Vienna, Austria). The isotopic composition of samples was normalized to a calibrated IVA soil
204 standard (1.65% C, $\delta^{13}\text{C} = -26.66$ ‰) measured under identical conditions every 10 samples.

205 The stable isotope ratio values are given in the delta notation with respect to the standard Vienna
206 Pee Dee Belemnite (VPDB).

207 *2.11. GC-c-IRMS*

208 GC-c-IRMS measurements were carried out with a Trace GC Ultra gas chromatograph
209 equipped with an on-column injector, Agilent HP5-MS column (30 m x 0.25 mm i.d. x 0.1 µm
210 film thickness), GC Isolink II conversion unit, comprising a combustion oven (at 1000 °C), a
211 ConFlo IV interface system and a Delta V Plus mass spectrometer (Thermo Scientific). The
212 temperature program was: 80 °C – 310 °C (4°C/min) – isothermal at 310 °C (40 min). Each
213 analysis was repeated 3x and the mean standard deviation was calculated.

214 Before and after each triplicate, the carbon isotopic composition of a certified *n*-alkane mixture
215 (Type A5; Arndt Schimmelmann, Department of Geological Sciences, Biogeochemical
216 Laboratories, Indiana University, USA) was measured and used for calibration. The stability of
217 the measurements was checked using pulses of reference CO₂ prior (5 pulses) and after (3
218 pulses) each run. The data were analyzed using Isodat 3.0 software. For correction of the δ¹³C
219 values of derivatized lipids, see Section 2.4.

220 *2.12. Radiocarbon dating from AMS*

221 The ¹⁴C measurements were performed in the Laboratory of Ion Beam Physics (ETH of Zurich,
222 Switzerland) using the MICADAS AMS equipment (Synal et al., 2007) which possess a gas
223 handling system adapted for very small volumes (µg level; Ruff et al., 2010; Wacker et al.,
224 2010a,b). Samples containing > 100 µg C as well as phthalic acid (blank analyses) were
225 graphitized using the automated AGE system (Wacker et al., 2010a). Small samples (i.e isolated
226 miliacin) were oxidized (combusted) and the resulting CO₂ was transferred to quartz tubes for
227 analysis in the ion source (Ruff et al., 2007, 2010). A standard (oxalic acid II) was prepared and

228 measured together with the samples. $^{14}\text{C}/^{12}\text{C}$ and $^{13}\text{C}/^{12}\text{C}$ isotopic ratios were determined using
229 the MICADAS dating system (Synal et al., 2007; Wacker et al., 2010b). Measured $^{14}\text{C}/^{12}\text{C}$
230 values of samples were corrected for the blanks and isotopic fractionation ($\delta^{13}\text{C}$, determined
231 with graphite or CO_2), ^{14}C age values were calculated following Stuiver and Polach (1977).
232 Calibrated ages were established using the atmospheric calibration curve IntCal13 reported by
233 Reimer et al. (2013).

234 **3. Results and discussion**

235 *3.1. Evidence from miliacin for millet cropping*

236 Overall, at the Obernai site, the lipid signatures from the soils sampled within the silos (e.g. silo
237 1330; Fig. 2a), in the loess substrate and in the Gaulish pit (structure 1001; Fig. 2b,c) showed
238 the same global lipid assemblage, comprising *n*-alkyl lipids (alkanes, FAs and alcohols), sterols
239 and triterpenoids. *n*-Alcohols in the C_{22} - C_{32} range were predominant and showed an even
240 carbon number predominance, with C_{26} prevailing. Even numbered *n*-acids from higher plants
241 in the C_{20} - C_{32} range were systematically dominated by C_{24} - C_{30} , whereas *n*-alkanes with an
242 odd/even predominance were generally dominated by C_{29} and C_{31} . These straight chain
243 distributions reflect the contribution from terrestrial plants, possibly from grassland plants (e.g.
244 van Bergen et al., 1997; Trendel et al., 2010). Within most silos and in the Gaulish pit and
245 latrines (Fig. 2), C_{29} sterols (**1-3**; numbers refer to structures in Appendix A), as well as a series
246 of pentacyclic triterpenoids comprising glutinone (**4**), taraxerol (**5**), germanicol (**6**), lupeol (**7**),
247 friedelin (**8**), α -amyrin (**9**), bauerenyl acetate (**10**) and betulin (**11**), again point toward a
248 predominant higher plant contribution (e.g. Lavrieux et al., 2011; Schnell et al., 2014). Closer
249 examination of the triterpene assemblage from the soil samples filling some of the
250 archaeological structures (silos, Gaulish pit, latrines; Fig. 2) revealed the presence of an early
251 eluting triterpenoid identified as olean-18-en-3 β -ol methyl ether, i.e. miliacin- (**12**), based on

252 its mass spectrum (Jacob et al., 2008b). This rather rare compound belongs to the series of
253 pentacyclic triterpene methyl ethers (PTMEs) generally restricted to Gramineae (Ohmoto et al.,
254 1970; Jacob et al. 2005). The presence of a methoxy group at C-3 has been shown to give a
255 high resistance of PTMEs towards biodegradation, which additionally display antibacterial and
256 antifungal properties (Jacob et al., 2005). This enhanced resistance allowed miliacin to be
257 detected within palaeosoils investigated in an archaeological context (Motuzaite-Matuzeviciute
258 et al., 2016). As shown by Jacob et al. (2008b) and Bossard et al. (2013), miliacin is closely
259 linked to its specific precursor organism, *Panicum miliaceum* (broomcorn millet or common
260 millet), where it occurs in the different plant compartments (seeds, stems, ears, leaves, roots),
261 but is more concentrated by far in the grains. In contrast, it is completely absent among the
262 triterpenoids from another widespread millet species, *Setaria italica* (foxtail millet; Bossard et
263 al., 2013), allowing a distinction to be made at the molecular level between these two cereal
264 species. Given this specificity, detection of millet within most of the soils filling the silos at the
265 Obernai site is of particular interest. Its presence indeed suggests that common millet was
266 cultivated at the site, with the possibility that this cereal was stored within the silos (see below).
267 Furthermore, other minor triterpenoids also biosynthesized by the broomcorn millet - although
268 in much lower quantity than miliacin - comprising isosawamilletin (**13**) - another PTME -
269 together with germanicol (**6**) and friedelin (**8**) (Bossard et al., 2013), co-occur with miliacin in
270 the archaeological soils (Fig. 2a). Another piece of evidence confirming that millet was present
271 at the site was recently obtained from carpological studies (Durand, unpublished results) which
272 led to the identification of a few carbonized and mineralized grains of *P. miliaceum* within some
273 silos. However, the low number of grains (< 10) generally identified in the different silos -
274 except in the case of silo 1763 in which 172 remnants of carbonized grains were identified in
275 the lower archaeological layer (AL2; Fig. 1) - did not allow a correlation to be established
276 between the occurrence of cereal remains in the silos and the presence of miliacin in the

277 corresponding soil samples. Some silos indeed showed a complete absence of millet grains, but
278 where miliacin was detected. Finally, stable carbon isotopic studies provided additional
279 evidence indicating that miliacin found at Obernai and common millet are closely linked. The
280 $\delta^{13}\text{C}$ value of miliacin was determined at ca. -21.5‰ (Table 2), in the same range as the values
281 reported for miliacin isolated from reference millet grains or extracted from soils (-21.5 to -
282 23.5‰; Jacob et al., 2008b). These values, around -20‰, are much less ^{13}C -depleted than those
283 generally determined in the case of lipids from temperate region plants which generally use the
284 C_3 carbon fixation pathway. This pathway leads to lipids having $\delta^{13}\text{C}$ values between -30 and -
285 40‰ (e.g. Collister et al., 1994; Chikaraishi et al., 2004; Bi et al., 2005; Chikaraishi and
286 Naraoka, 2006), whereas *P. miliaceum* has a C_4 metabolism (Jacob et al., 2008b). The
287 latter leads to the biosynthesis of biomass (including lipids) less ^{13}C -depleted (e.g., Chikaraishi
288 et al., 2004; Bi et al., 2005; Chikaraishi and Naraoka, 2007; Mendez-Millan et al., 2014). In
289 the case of miliacin detected at the Obernai site, the $\delta^{13}\text{C}$ composition is in good agreement
290 with a C_4 plant source, confirming that it most likely originates from *P. miliaceum* (Jacob,
291 2008b).

292 3.2. Role of archaeological pits as traps for lipid biomarkers

293 As determined above, based on molecular, isotopic and carpologic grounds, millet seemed to
294 be present and likely cultivated as a cereal at the site of Obernai. Given the presence of miliacin
295 in most of the silos, it can be proposed, at first glance, that millet was stored in the silos, at least
296 in those dating from the Middle Bronze Age (silo 1763) up to the Second Iron Age (silos 1330,
297 1508, 1513 and 1521). If this were the case, miliacin would represent the molecular remains of
298 the grains originally stored in the silos and recovered for consumption. One would then expect
299 to find the highest concentration of miliacin at the bottom of the silos, and eventually trace
300 amounts (if any) in the AL from the middle and upper soil filling. To test this hypothesis,

301 quantitative analysis of miliacin by way of GC-FID was performed on AL from silo soil
302 samples at different filling depths. Except in the case of silo 1817 (Neolithic), in which no
303 miliacin was found (see discussion below), all LE from the silo soils contained miliacin in high
304 relative amount (Fig. 3), whereas it was absent - or present at trace level - in the loess substrate.
305 This strongly suggests that the presence of miliacin is closely associated with the silos. However,
306 miliacin concentration profiles within the different silos showed high variability. In silos 1513,
307 1508 and 1763, the highest concentration was at the bottom, whereas the opposite was the case
308 for silos 1521 and 1781 (Fig. 3). There was also an intermediate situation, with the highest
309 concentration in the middle sampling levels (silo 1330). These unexpected concentration
310 profiles suggested that miliacin (and hence, millet cultivation) was closely associated with the
311 surface horizons of surrounding soils that filled the silos rather than indicating that millet was
312 the cereal stored in the silos. This apparent disjunction between the function of an
313 archaeological structure (cereal storage in silos, in the present case) and its lipid content is also
314 illustrated by the presence of miliacin in the LE from a Gaulish pit sample - an archaeological
315 structure not related to any agricultural practices - from the same archaeological site (Fig. 2c).

316 In conclusion, with respect to the presence of millet at this archaeological site, we propose that
317 the silos, as well as the Gaulish pit, played the role of "archaeological traps", in the sense that
318 these structures, once abandoned, were filled quickly (intentionally or not) with surrounding
319 soil horizon(s), the latter having been "sealed" within these structures and having kept the record
320 of past agricultural practices. This conclusion would be in line with the concept of "pedological
321 trap" recently developed (e.g., Ertlen et al., 2013; Lauer et al., 2013).

322 *3.3. Evidence from ¹⁴C dating for millet cropping dating back to the protohistoric and antic*
323 *periods*

324 In order to constrain the timing of millet cultivation/consumption at the Obernai site, ^{14}C dating
325 measurements were undertaken on SOM, LE, isolated miliacin from a series of selected
326 archaeological samples and millet grains from silo 1763 (Féliu, 2017). Among them, two silos
327 were chosen as representative of distinct periods of occupation of the site, based on ceramology
328 dating (Table 1). Silo 1763 represents the oldest structure in which miliacin was detected (see
329 above) and is dated from the Late Bronze Age (950-800 BC, corresponding to ca. 2900-2750
330 yr BP), whereas silo 1330 was selected to cover the Second Iron Age (400-260 BC, ca. 2350-
331 2210 yr BP). A third, more recent structure (4564) from the site - not discussed in detail here -
332 corresponding to latrines from the Roman Gaul period (end of the 2nd - beginning of the 3rd
333 century AD, ca. 1750-1800 yr BP), was also shown to contain exceptionally high amounts of
334 miliacin (Fig. 2d), which was isolated from AL8 and ^{14}C dated (Table 3).

335 Given the low amount of OM generally in these loess soil samples ($\text{TOC} < 0.1\%$), radiocarbon
336 dating measurements were carried out using the AMS instrument MICADAS. Pure miliacin
337 from silos 1330 and 1763 and Roman-Gaul latrines 4564 was obtained from the LE from AL
338 (Table 3) using a combination of separation steps involving silica gel liquid chromatography,
339 reversed phase HPLC and thin layer chromatography (Section 2.6). Using our isolation protocol,
340 miliacin was recovered in the range of 15 μg to 300 μg , with a purity between 80% and > 90%
341 (determined from GC, Fig. S2 in Supplementary data), the co-occurring compounds comprising
342 mainly olean-13(18)-en-3 β -ol methyl ether (isosawamilletin, **13**) another triterpenoid from
343 millet (Bossard et al., 2013). Despite the rather low quantity isolated in most cases, ^{14}C age
344 values could be determined as ^{14}C age BP and calibrated ^{14}C age BC (Table 3 and Table S2 for
345 the complete data set).

346 In the case of silo 1330 (Second Iron Age), the ^{14}C age of SOM from a soil sample collected
347 within AL6 located at mid-depth of the silo (Fig. 1) was measured at 3042 yr BP, slightly older
348 than the lipid extract from the same soil sample by ca. 200 yr (Fig. 4 and Table 3). The age of

349 miliacin isolated from this soil sample, as well as that from miliacin from two AL located at the
350 bottom and top of the same silo (AL2 and AL10) showed globally the same age of ca. 3000-
351 3200 yr BP (Fig. 4 and Table 3). Interestingly, the ¹⁴C age of miliacin from the second silo (silo
352 1763, Late Bronze Age) was similar to that of miliacin from silo 1330 (Second Iron Age). These
353 values, all close to that determined for SOM from AL6 (silo 1330) and AL2 (silo 1763), are
354 clearly older than the age of silo structure 1330 as determined from ceramology (ca. 400-260
355 BC), indicating that miliacin (and hence, millet) was not contemporaneous with the use of the
356 silos for cereal storage. In fact, in all cases, the age of miliacin corresponds to the Bronze Age,
357 between 3994 and 2776 yr BP, whereas silos 1763 and 1330 were probably in use between 950-
358 800 BC (end of the Bronze Age) and 400-260 BC (second Iron Age), respectively.

359 The age difference between the archaeological structure and miliacin was particularly marked
360 with silo 1330, with a difference > 600 yr. Moreover, the similarity in the age of miliacin from
361 the three archaeological layers sampled within silo 1330 indicates that the structure was filled
362 quickly with the same soil after abandoning, whatever the archaeological layers considered.
363 The situation was different with silo 1763, for which miliacin had almost the same (or slightly
364 older) age (Fig. 4) than the silo dated from the end of the Bronze Age, a period during which
365 the cereal was cultivated and likely stored in silos. A confirmation that millet was consumed at
366 this time period was indeed attested to from the presence, at the bottom of the structure (AL2;
367 Fig. 1), of a few carbonized millet grains, dated at ca. 2800 yr BP (2880-2762 cal yr BP for 2σ;
368 Féliu, 2017).

369 In conclusion, based on quantitative and ¹⁴C dating measurements, it appears that the presence
370 of miliacin cannot be interpreted as evidence of cereal storage in the silo structures, contrary to
371 what was initially envisaged, but rather reflects the existence of millet cultivation predating the
372 digging of the silos, except silos 1817 (Neolithic, predating millet cropping), and 1763 (Late
373 Bronze Age, contemporaneous with millet cropping). Once abandoned, the emptied silos were

374 filled rapidly with the surface horizons of surrounding soils and thus acted as "pedological
375 traps" for the soils that were used for millet cultivation ca. 3 kyr BP ago, sealing and preserving
376 their SOM content, including lipids. A search for biomarkers indicative of the contribution of
377 other types of cereals, such as wheat or barley, such as e.g., alkyl resorcinols (Appendix A;
378 Hengtrakul et al., 1991; Seitz, 1992; Ross et al., 2003, 2004), was unsuccessful. Indeed, there
379 was no evidence for such an occurrence from the lipid extracts, whereas these cereals could be
380 detected in the form of carbonized grains from carpological studies, found within some silos
381 dated from the Late Bronze Age and La Tène (Durand, unpublished results). The absence of
382 alkyl resorcinols could be due to a low preservation potential, as these compounds have been
383 shown to be easily degraded upon processes like fermentation or heating (Winata and Lorenz,
384 1997), possibly explaining why they have never been reported in archaeological studies.

385 The case of the Roman Gaul latrine structure 4564 is sharply different from that of the silos,
386 since an almost identical ^{14}C age was obtained for the LE, SOM and miliacin, this age being
387 also in good agreement with that of the structure itself, as determined independently from
388 ceramology. In this case, miliacin consumption (and/or millet cultivation) could be
389 unambiguously correlated with the Roman Gaul period, and was confirmed from carpological
390 studies showing the presence - although in very low amount - of mineralized millet grains
391 within the latrine structure (Durand, unpublished results).

392 *3.4. C_4 vs. C_3 plant input using the $\delta^{13}\text{C}$ record of n -alkyl lipids*

393 Several studies have used the C isotopic changes of SOM or lipids within soils to trace back C_3
394 vs. C_4 vegetation change and to study the turnover of SOM induced by the vegetation change
395 (e.g. Schwartz, 1991; Mariotti and Peterschmitt, 1994; Pessenda et al., 1998; Wiesenberg et al.,
396 2004; Quénéa et al., 2006; Mendez-Millan et al., 2014). In the present study, the stable carbon
397 isotopic composition of some straight chain lipids (n -alkanes, FAs and alcohols) from selected

398 soil samples was measured in order to determine if *n*-alkyl inputs from a C₄ plant (i.e., millet)
399 could be detected on the basis of these compounds. It appeared that the *n*-alkanes in the LE
400 from the soils filling silos 1330, 1781 and 1508 showed $\delta^{13}\text{C}$ values between -29.9 and -35.5‰
401 (Table 2) typical for C₃ plants (Bi et al., 2005; Mendez-Millan et al., 2014). The same held for
402 the FAs - when they could be measured - with $\delta^{13}\text{C}$ values ranging between -31.6‰ and -37.0‰
403 (Table 2). Similar values were obtained in the case of the *n*-alcohols ($\delta^{13}\text{C}$ between -31.4‰ and
404 -36.8‰; Table 2). These values highlight an almost pure C₃ isotopic signature. This was
405 confirmed by the $\delta^{13}\text{C}$ values of bulk SOM in selected samples (silo 1330 AL 2, 6, 10 and
406 Roman latrines 4564 AL 4, 8 9), which all fell in the rather narrow range of $-25.5\text{‰} \pm 1$,
407 corresponding to a C₃ source (e.g. Balesdent et al., 1987; Schwartz, 1991; Beniston et al., 2014;
408 Chikaraishi et al., 2004). Interestingly, the $\delta^{13}\text{C}$ values of SOM from the corresponding
409 surrounding soils gave the same isotopic signature, indicating that the C₄ contribution detected
410 at the molecular level (i.e. the presence of miliacin) could not be observed from bulk carbon
411 analysis.

412 For the lipids, the case of 1-dotriacontanol (*n*-C₃₂) was slightly different from the other straight
413 chain compounds, with a $\delta^{13}\text{C}$ value enriched in ¹³C by ca. 2-4‰ compared with the lower *n*-
414 alcohol C₂₆-C₃₀ homologues (Table 2), suggesting a mixed contribution from both C₃ and C₄
415 plants. This is the case in all the silo samples containing miliacin and, interestingly, not the case
416 for the loess substrate samples and for the Neolithic silo 1817, the sole silo devoid of evidence
417 for millet occurrence (Table 2). It turns out that our study of common millet highlighted the
418 particular abundance of 1-dotriacontanol in the leaves (Fig. S3 in Supplementary data) in
419 accordance with the literature (up to 80% of summed free *n*-alcohols according to Tulloch,
420 1982). Consequently it could be envisaged that part of this compound coming from millet was
421 preserved, whereas the C₄ $\delta^{13}\text{C}$ signature of the other straight chain lipids, originally less

422 abundant in millet, was progressively replaced with the C₃ δ¹³C signature of the succeeding
423 vegetation.

424 In conclusion, most of the soil lipids, as well as the SOM showed a clear C₃ plant signal, most
425 likely reflecting the contribution from Gramineae. In the soils filling the silos, this predominant
426 contribution was accompanied by a small signal from a C₄ plant source, mainly highlighted by
427 the triterpenoid miliacin and its δ¹³C signature, but also, to a minor extent, by the δ¹³C signal
428 of 1-dotriacontanol.

429 *3.5. Extent of millet cropping inferred from stable carbon isotopes and ¹⁴C dating*

430 The low contribution of lipids from millet to the *n*-alkyl lipid pool, as inferred from stable
431 carbon isotopes, could be explained on one hand by a limited contribution from millet biomass
432 to the SOM vs. that of C₃ plants (i.e. limited millet cropping). Such a situation might be
433 encountered if millet cropping were not quantitatively important in terms of surface cultivation
434 or of duration of cultivation. In this case, the molecular signature of *n*-alkyl lipids from millet
435 would be diluted with that from the predominating surrounding C₃ vegetation (natural or
436 cultivated in parallel with millet). Furthermore, the predominant C₃ vegetation that either
437 preceded or succeeded millet cropping might also be responsible for the observed isotopic
438 signature for the filling soils, a signature which was "sealed" within the silos after they were
439 abandoned.

440 In addition, irrespective of the importance of the initial contribution of millet biomass to SOM,
441 the stable carbon isotopic composition of straight chain lipids and miliacin within silo 1330 -
442 but probably also the other silos from the Iron Age - could be interpreted in terms of lipid
443 turnover associated with a vegetation change from a mixed C₃+C₄ vegetation to a pure C₃
444 vegetation cover in the light of the radiocarbon dating results. Indeed, there is a time difference
445 of several centuries between the age of miliacin (Bronze Age) and that of the silo (Iron Age) in

446 which miliacin was detected (Table 3). This difference implies that, following millet cultivation,
447 the soils underwent vegetation change before being trapped within the abandoned silo, this
448 vegetation being made up of C₃ plants (e.g. pasture or cereals like wheat, barley, ...) based on
449 the $\delta^{13}\text{C}$ values of the lipids and SOM (see above). The period during which the C₄ plus C₃
450 vegetation lasted cannot be determined, the radiocarbon age of miliacin corresponding to an
451 average age resulting from more ancient and more recent contributions, with unknown limits
452 for the beginning and end of this cereal crop. The progressive replacement of millet cropping
453 resulted in the “dilution” of the millet molecular and isotopic signatures by those of the newly
454 occupying C₃ plants. For each molecular structure considered, the replacement rate would
455 depend on one hand on the relative contribution of millet origin vs. C₃ plants and on the other
456 hand on the relative stability of each structure considered. For instance, miliacin, which has a
457 particularly stable structure due to the presence of the methoxy group at C-3, and which is not
458 biosynthesized by C₃ plants native to temperate regions (Bossard et al., 2013), would keep its
459 typical C₄ isotopic signature (i.e., no isotopic "dilution" given the absence of miliacin inputs
460 from a C₃ source). By contrast, in the case of straight chain lipids, the progressive degradation
461 of the C₄ source, together with the constant new input from straight chain lipids of C₃ origin,
462 would result in the progressive "dilution" of the molecular and isotopic signature of lipids of C₄
463 origin. Such a situation has some analogy with that reported by Mendez-Millan et al. (2014),
464 who investigated the evolution of the stable carbon isotopic composition of lipids from a soil
465 during a 30 yr chronosequence during which savannah (in this case dominated by C₄ plants)
466 was replaced by an eucalyptus plantation (a C₃ plant). Here, the molecular and isotopic
467 fingerprints from triterpenoid methyl ethers from savannah vegetation was only little affected
468 by the vegetation change, whereas the isotopic composition of the straight chain lipids was
469 progressively shifted towards more negative $\delta^{13}\text{C}$ values, as expected from such a vegetation
470 change. Based on stable carbon isotopes, it was notably determined that > 30 % of the *n*-C₂₇

471 alkane originated from the newly installed C₃ plant source after 30 yr of eucalyptus coverage.
472 Conversely, in the case of a vegetation change from C₃ (wheat) to C₄ (maize) plants, the δ¹³C
473 values from soil constituents move towards less ¹³C-depleted values (Lichtfouse et al., 1995).

474 *3.6. Timing of millet cropping at the site of Obernai*

475 Contrary to the protohistoric silos described above, the filling soil from silo 1817 (Mid-
476 Neolithic, 4600-4300 BC) was devoid of miliacin, suggesting that millet was not cultivated at
477 that time period. By contrast, structure 4564 (Roman Gaul period), corresponding to latrines,
478 contained very high amounts of miliacin, having the same radiocarbon age (Table 3) as that of
479 the structure. In this case, it seems that millet was consumed and likely cultivated locally at this
480 period, in agreement with the literature (Reddé et al., 2005; Vandorpe and Wick, 2015).

481 The radiocarbon measurements allowed establishing that millet was cultivated during the
482 Bronze Age and the Roman Gaul period at least, which makes sense in the light of the historical
483 and archaeological data for the history of this cereal. In Alsace (NE France), where agricultural
484 practices can be traced back down to ca. 4500 BC (Ruas and Marinval, 1991), very few
485 archaeological studies report the detection of carpologic remains from millet during
486 Protohistoric times, with only limited mention of its cultivation and consumption during the
487 Iron Age (Wiethold, 2000; Bataille, 2014; Goude et al., 2015). This is also the case during the
488 Roman period (Reddé et al., 2005; Vandorpe and Jacomet, 2011; Vandorpe and Wick, 2015),
489 and the present study of the site of Obernai brings complementary information regarding millet
490 cropping at the local and national scale during this period.

491 From an archaeological viewpoint, millet finds its origin in China where it was domesticated
492 since the VIIth millenium BC (Laporte, 2001; Fuller, 2007; Lu et al., 2009). For instance, it has
493 been recently shown that, in China, millet grains were fermented with other cereal grains to
494 prepare beer (Wang et al., 2016). Later, millet culture began to spread through Asia and the

495 Middle East, before reaching Central Europe at the end of the Neolithic (Hunt et al., 2008).
496 However, whereas in France wheat and barley were cultivated since the Neolithic (Ruas and
497 Marinval, 1991; Martin et al., 2007), it is only during the Protohistoric times that cereal species
498 became more diversified, and the first evidence from carpology of *P. miliaceum* culture can be
499 dated back from the Early Bronze Age (ca. 1700 BC; Marinval, 1992, 1995). The Bronze and
500 Iron Ages are clearly associated with intensified cultivation of this cereal in France. Millet
501 grains were stored with their protecting envelopes (husks) for better preservation within the
502 buried silos -like those found at Obernai - or in vases (Marinval, 1995). Following the conquest
503 of Gaul, the Romans imported new varieties of cereals, mainly from the Mediterranean Basin,
504 resulting in enhanced diversification of foodstuffs (Cabanis et al., 2015). This resulted in the
505 progressive replacement of millet by other cereal species (e.g. rye), millet having been
506 completely abandoned at the beginning of the Middle Age (Jacob et al., 2008a).
507 Notwithstanding this, cultivation of millet lasted in Obernai at least until the end of the 2nd
508 century AD – beginning of the 3rd century AD.

509 **4. Conclusions**

510 The archaeological site of Obernai (NE France) has provided the opportunity to investigate past
511 local agricultural practices following unearthing of a series of protohistoric silos from the
512 Bronze and Iron Ages. GC-MS analysis of the lipids preserved in the soils filling the silos has
513 shown the presence of miliacin, a triterpenoid methyl ether strictly associated with the cereal *P.*
514 *miliaceum* (broomcorn millet). However, the *n*-alkyl lipid assemblage co-detected with miliacin
515 was associated to C3-plant sources for which no specific lipids were identified. The absence of
516 a C4 isotopic signature for the *n*-alkyl lipids was probably caused by the turnover of the SOM
517 induced by a change of vegetation cover. Radiocarbon dating of miliacin, SOM and lipid
518 extracts indicated that millet cropping was established during the Bronze Age and during the
519 Roman Gaul period. The study represents the first evidence of millet cultivation in Alsace

520 dating back to the Bronze Age, bringing a new perspective regarding agricultural practices and
521 past diet in the East of France.

522 This study has also highlighted the potential of hollow archaeological structures like pits and
523 silos to constitute “pedological traps” able to trap and archive soils surrounding the
524 archaeological structures during their period of use or after abandoning. These filling soils have
525 kept the molecular signatures from the various types of vegetation cover which developed on
526 the surface soil horizons prior to being trapped in the structures, thereby potentially providing
527 information about the past environment of archaeological sites in terms of vegetation cover or
528 agricultural practices. This is notably the case for miliacin found in silos from the Iron Age in
529 Obernai, attesting to millet cropping during the Bronze Age. The dating of lipids related to a
530 given biological source and of specific biomarkers, together with their stable carbon isotopic
531 composition, appeared to be critical for relating the molecular signatures to a specific
532 chronological/historical context.

533 **Acknowledgements**

534 The INRAP (Institut National de Recherches Archéologiques Préventives) is thanked for
535 providing the soil samples from the archaeological structures from Obernai, as is J. Jacob
536 (Institut des Sciences de la Terre, Université d’Orléans, France) for providing an authentic
537 miliacin standard and fresh material from broomcorn millet, and I. Antheaume and V. Grossi
538 (University Claude Bernard Lyon I) for the bulk $\delta^{13}\text{C}$ analysis of SOM. B.C. thanks the French
539 Ministère de l’Enseignement Supérieur et de la Recherche for a doctoral fellowship. Part of the
540 radiocarbon data was financed through the project IDEX 2015 ARCHEOSOL (University of
541 Strasbourg). L. Wacker is thanked for support via GIS AMS analysis at ETH Zurich.

542 **References**

543 Balesdent, J., Wagner, G.H., Mariotti, A., 1988. Soil organic matter turnover in long-term field
544 experiments as revealed by carbon-13 natural abundance. Soil Science Society of America
545 Journal 52, 118-124.

546 Ballentine, D.C., Macko, S.A., Turekian, V.C., 1998. Variability of stable carbon isotopic
547 compositions in individual fatty acids from combustion of C₄ and C₃ plants: implications for
548 biomass burning. Chemical Geology 152, 151-161.

549 Bataille, G., 2014. Eckbolsheim, Bas-Rhin. Parc d'activité – Les fouilles archéologiques
550 préalables à la construction des parkings et zones annexes du Zénith. Report, Vol. 2, INRAP
551 Grand Est Sud, Strasbourg.

552 Bi, X., Sheng, G., Liu, X., Li, C., Fu, J., 2005. Molecular and carbon and hydrogen isotopic
553 composition of *n*-alkanes in plant leaf waxes. Organic Geochemistry 36, 1405-1417.

554 Birkholz, A., Smittenberg, R.H., Hajdas, I., Wacker, L., Bernasconi, S.M., 2013. Isolation and
555 compound specific radiocarbon dating of terrigenous branched glycerol dialkyl glycerol
556 tetraethers (brGDGTs). Organic Geochemistry 60, 9-19.

557 Boëda, E., Connan, J., Dessort, D., Muhsen, S., Mercier, N., Valladas, H., Tisnérat, N., 1996.
558 Bitumen as a hafting material on Middle Palaeolithic artefacts. Nature 380, 336-338.

559 Bossard, N., Jacob, J., Le Milbeau, C., Sauze, J., Terwilliger, V., Poissonnier, B., Vergès, E.,
560 2013. Distribution of miliacin (olean-18-en-3 β -ol methyl ether) and related compounds in
561 broomcorn millet (*Panicum miliaceum*) and other reputed sources: Implications for the use of
562 sedimentary miliacin as a tracer of millet. Organic Geochemistry 63, 48-55.

563 Buckley, S.A., Evershed, R.P., 2001. Organic chemistry of embalming agents in Pharaonic and
564 Graeco-Roman mummies. Nature 413, 837-841.

565 Bull, I.D., Simpson, I.A., Dockrill, S.J., Evershed, R.P., 1999. Organic geochemical evidence
566 for the origin of ancient anthropogenic soil deposits at Tofts Ness, Sanday, Orkney. *Organic*
567 *Geochemistry* 30, 535-556.

568 Bull I.D., Betancourt P.P., Evershed R.P., 2001. An organic geochemical investigation of the
569 practice of manuring at a Minoan site on Pseira Island, Crete. *Geoarchaeology* 16, 223-242.

570 Cabanis, M., Deberge, Y., Bouby, L., Hajnalová, M., Marinval, P., Mennessier-Jouannet, C.,
571 Vermeulen, C., Vernet, G., 2015. Changes in crop cultivation during the last five centuries
572 before the Roman conquest: archaeobotanical investigation in the Clermont-Ferrand basin,
573 Massif Central, France. *Archaeological and Anthropological Sciences*, 1-16.

574 Charrié-Duhaut, A., Connan, J., Rouquette, N., Adam, P., Barbotin, C., de Rozières, M.-F.,
575 Tchaplá, A., Albrecht, P., 2007. The canopic jars of Rameses II: real use revealed by molecular
576 study of organic residues. *Journal of Archaeological Science* 34, 957-967.

577 Charters, S., Evershed, R.P., Blinkhorn, P.W., Denham, V., 1995. Evidence for the mixing of
578 fats and waxes in archaeological ceramics. *Archaeometry* 37, 113-127.

579 Chikaraishi, Y., Naraoka, H., Poulson, S.R., 2004. Hydrogen and carbon isotopic fractionations
580 of lipid biosynthesis among terrestrial (C₃, C₄ and CAM) and aquatic plants. *Phytochemistry*
581 65, 1369-1381.

582 Chikaraishi, Y., Naraoka, H., 2006. Carbon and hydrogen isotope variation of plant biomarkers
583 in a plant-soil system. *Chemical Geology* 231, 190-202.

584 Chikaraishi, Y., Naraoka, H., 2007. $\delta^{13}\text{C}$ and δD relationships among three *n*-alkyl compound
585 classes (*n*-alkanoic acid, *n*-alkane and *n*-alkanol) of terrestrial higher plants. *Organic*
586 *Geochemistry* 38, 198-215.

587 Christl, M., Vockenhuber, C., Kubik, P.W., Wacker, L., Lachner, J., Alfimov, V., Synal, H.-
588 A., 2013. The ETH Zurich AMS facilities: performance parameters and reference materials.
589 Nuclear Instruments and Methods in Physics Research B 294, 29-38.

590 Collister, J.W., Rieley, G., Stern, B., Eglinton, G., Fry, B., 1994. Compound-specific $\delta^{13}\text{C}$
591 analyses of leaf lipids from plants with differing carbon dioxide metabolisms. Organic
592 Geochemistry 21, 619-627.

593 Condamin, J., Formenti, F., Metais, M.O., Michel, M., Blond, P., 1976. The application of gas
594 chromatography to the tracing of oil in ancient amphorae. Archaeometry 18, 195-201.

595 Connan, J., 1999. Use and trade of bitumen in antiquity and prehistory: molecular archaeology
596 reveals secrets of past civilizations. Philosophical Transactions of the Royal Society B:
597 Biological Sciences 354, 33-50.

598 Courel, B., Schaeffer, P., Adam, P., Ertlen, D., Schwartz, D., Bernasconi, S.M., Hajdas, I., 2015.
599 Analyse, isolement et datation au ^{14}C de lipides dans les sols. L'exemple des tétraéthers de
600 diglycérol. In : Poulenard, J., Rossi, M. (Eds.), Sols et Matières Organiques: Mémoires et
601 Héritages. EDYTEM. Université de Savoie, Le Bourget-du Lac, pp 57-68.

602 Egli, M., Gristina, L., Wiesenberg, G.L.B., Civantos, J.M.M., Rotolo, A., Novara A., Brandová,
603 D., Raimondi, S., 2013. From pedologic indications to archaeological reconstruction:
604 deciphering land use in the Islamic period in the Baida district (north-western Sicily). Journal
605 of Archaeological Science 40, 2670-2685.

606 Eglinton, G., Logan, G.A., 1991. Molecular preservation. Philosophical Transactions of the
607 Royal Society B: Biological Sciences 333, 315-328.

608 Ertlen, D., Féliu, C., Michler, M., Schneider-Schwien, N., Schneikert, F., Thomas, Y., Braguier,
609 S., Minni, D., Veber, C., 2013. Les structures de type « fente » dans le Kochersberg (Alsace):

610 un programme de recherche en cours de développement. *Revue archéologique de l'Est*,
611 Supplément 33, 213-227.

612 Ertlen, D., Schwartz, D., Brunet, D., Trendel, J.-M., Adam, P., Schaeffer, P., 2015. Qualitative
613 near infrared spectroscopy, a new tool to recognize past vegetation signature in soil organic
614 matter. *Soil Biology & Biochemistry* 82, 127-134.

615 Evershed, R.P., Heron, C., Goad, L.J., 1991. Epicuticular wax components preserved in
616 potsherds as chemical indicators of leafy vegetables in ancient diets. *Antiquity* 65, 540-544.

617 Fuller, D.Q., 2007. Contrasting patterns in crop domestication and domestication rates: recent
618 archaeobotanical insights from the Old World. *Annals of Botany* 100, 903-924.

619 Gierga, M., Hajdas, I., van Raden, U.J., Gilli, A., Wacker, L., Sturm, M., Bernasconi, S.M.,
620 Smittenberg, R.H., 2016. Long-stored soil carbon released by prehistoric land use: evidence
621 from compound-specific radiocarbon analysis on Soppensee lake sediments. *Quaternary*
622 *Science Reviews* 144, 123-131.

623 Goude, G., Balasescu, A., Réveillas, H., Thomas, Y., Lefranc, P., 2015. Diet variability and
624 stable isotope analyses: looking for variables within the Late Neolithic and Iron Age human
625 groups from Gougenheim site and surrounding areas (Alsace, France). *International Journal of*
626 *Osteoarchaeology* 25, 988-996.

627 Hengtrakul, P., Lorenz, K., Mathias, M., 1991. Alkylresorcinol homologs in cereal grains.
628 *Journal of Food Composition and Analysis* 4, 52-57.

629 Huang, Y., Bol, R., Harkness, D.D., Ineson, P., Eglinton, G., 1996. Post-glacial variations in
630 distributions, ¹³C and ¹⁴C contents of aliphatic hydrocarbons and bulk organic matter in three
631 types of British acid upland soils. *Organic Geochemistry* 24, 273-287.

632 Hunt, H.V., Linden, M.V., Liu, X., Motuzaite-Matuzeviciute, G., Colledge, S., Jones, M.K.,
633 2008. Millets across Eurasia: chronology and context of early records of the genera *Panicum*
634 and *Setaria* from archaeological sites in the Old World. *Vegetation History and Archaeobotany*
635 17 (Suppl 1), S5-S18.

636 Ingalls, A.E., Pearson, A., 2005. Ten years of compound-specific radiocarbon analysis.
637 *Oceanography* 18, 18-31.

638 Jacob, J., Disnar, J.-R., Boussafir, M., Albuquerque, A.L.S., Sifeddine, A., Turcq, B., 2005.
639 Pentacyclic triterpene methyl ethers in recent lacustrine sediments (Lagoa do Caçó, Brazil).
640 *Organic Geochemistry* 36, 449-461.

641 Jacob, J., Disnar, J.-R., Arnaud, F., Chapron, E., Debret, M., Lallier-Vergès, E., Desmet, M.,
642 Revel-Rolland, M., 2008a. Millet cultivation history in the French Alps as evidenced by a
643 sedimentary molecule. *Journal of Archaeological Science* 35, 814-820.

644 Jacob, J., Disnar, J.-R., Bardoux, G., 2008b. Carbon isotope evidence for sedimentary miliacin
645 as a tracer of *Panicum miliaceum* (broomcorn millet) in the sediments of Lake le Bourget
646 (French Alps). *Organic Geochemistry* 39, 1077-1080.

647 Koch, P.L., Fogel, M.L., Tuross, N., 1994. Tracing the diets of fossil animals using stable
648 isotopes. In: Lajtha, K., Michener, R.H. (Eds.), *Stable Isotopes in Ecology and Environmental*
649 *Science*, 1st Edition. Blackwell Scientific Publications, pp. 63-92.

650 Kögel-Knabner, I., 2002. The macromolecular organic composition of plant and microbial
651 residues as inputs to soil organic matter. *Soil Biology & Biochemistry* 34, 139-162.

652 Laporte, L., 2001. Guilaine, (J.), Dir. – Premiers paysans du monde, naissance des agricultures.
653 Séminaire du Collège de France. *Revue Archéologique de l'Ouest* 18, 240-242.

654 Lavrieux, M., Jacob, J., Le Milbeau, C., Zocatelli, R., Masuda, K., Bréheret, J.-G., Disnar, J.-
655 R., 2011. Occurrence of triterpenyl acetates in soil and their potential as chemotaxonomical
656 markers of *Asteraceae*. *Organic Geochemistry* 42, 1315-1323.

657 Lavrieux, M., Bréheret, J.-G., Disnar, J.-R., Jacob, J., Le Milbeau, C., Zocatelli, R., 2012.
658 Preservation of an ancient grassland biomarker signature in a forest soil from the French Massif
659 Central. *Organic Geochemistry* 51, 1-10.

660 Lavrieux, M., Jacob, J., Disnar, J.-R., Bréheret, J.-G., Le Milbeau, C., Miras, Y., Andrieu-Ponel,
661 V., 2013. Sedimentary cannabinoles track the history of hemp retting. *Geology* 41, 751-754.

662 Lichtfouse, E., Dou, S., Houot, S., Barriuso, E., 1995. Isotope evidence for soil organic carbon
663 pools with distinct turnover rate -II. Humic substances. *Organic Geochemistry* 23, 845-847.

664 Lichtfouse, E., Bardoux, G., Mariotti, A., Balesdent, J., Ballentine, D.C., Macko, S.A., 1997.
665 Molecular, ^{13}C , and ^{14}C evidence for the allochthonous and ancient origin of C_{16} - C_{18} *n*-alkanes
666 in modern soils. *Geochimica et Cosmochimica Acta* 61, 1891-1898.

667 Lorenz, K., Lal, R., Preston, C.M., Nierop, K.G.J., 2007. Strengthening the soil organic carbon
668 pool by increasing contributions from recalcitrant aliphatic bio(macro)molecules. *Geoderma*
669 142, 1-10.

670 Lu, H., Zhang J., Liu, K.-B., Wu, N., Li, Y., Zhou, K., Ye, M., Zhang, T., Zhang, H., Yang, X.,
671 Shen, L., Xu, D., Li, Q., 2009. Earliest domestication of common millet (*Panicum miliaceum*)
672 in East Asia extended to 10,000 years ago. *Proceedings of the National Academy of Sciences*
673 USA 106, 7367-7372.

674 Marinval, P., 1992. Archaeobotanical data on millets (*Panicum miliaceum* and *Setaria italica*)
675 in France. *Review of Palaeobotany and Palynology* 73, 259-270.

676 Marinval, P., 1995. Données carpologiques françaises sur les Millets (*Panicum miliaceum* L. et
677 *Setaria italica* (L.) Beauv.) de la Protohistoire au Moyen Age. In: Hörandner, E. (Ed.), Millet:
678 Actes du Congrès d'Aizenay, 18-19 août 1990. Peter Lang, Frankfurt, pp. 31-61.

679 Mariotti, A., Peterschmitt, E., 1994. Forest savanna ecotone dynamics in India as revealed by
680 carbon isotope ratios of soil organic matter. *Oecologia* 97, 475-480.

681 Martin, L., Thiébault, S., Rey, P.-J., 2007. Production et consommation végétale durant la
682 Préhistoire récente: études archéobotaniques de quelques sites des Alpes occidentales. In:
683 Boëtsch, G., Hubert, A (Eds.), Alimentation et Montagne, Actes de la 6^{ème} Université
684 Européenne d'Eté « Anthropologie des Populations Alpines », Gap, France. Editions des
685 Hautes-Alpes, Gap, pp. 11-24.

686 Mendez-Millan, M., Nguyen Tu, T.T., Balesdent, J., Derenne, S., Derrien, D., Egasse, C.,
687 Thongo M'Bou, A., Zeller, B., Hatté, C., 2014. Compound-specific ¹³C and ¹⁴C measurements
688 improve the understanding of soil organic matter dynamics. *Biogeochemistry* 118, 205-223.

689 Mollenhauer, G., Rethemeyer, J., 2009. Compound-specific radiocarbon analysis - analytical
690 challenges and applications. IOP Conferences Series: Earth and Environmental Science 5,
691 012006.

692 Motuzaitė-Matuzevičiūtė, G., Jacob, J., Telizhenko, S., Jones, M.K., 2016. Miliacin in
693 palaeosols from an Early Iron Age in Ukraine reveals *in situ* cultivation of broomcorn millet.
694 *Archaeological and Anthropological Sciences* 8, 43-50.

695 Ohmoto, T., Ikuse, M., Natori, S., 1970. Triterpenoids of the *Gramineae*. *Phytochemistry* 9,
696 2137-2148.

697 O'Leary, M.H., 1988. Carbon isotopes in photosynthesis. *BioScience* 38, 328-336.

698 Pessenda, L.C.R., Gomes, B.M., Aravena, R., Ribeiro, A.S., Boulet, R., Gouveia, S.E.M., 1998.
699 The carbon isotope record in soils along a forest-cerrado ecosystem transect: implications for
700 vegetation changes in the Rondonia state, southwestern Brazilian Amazon region. The
701 Holocene 8, 599-603.

702 Quéneá, K., Largeau, C., Derenne, S., Spaccini, R., Bardoux, G., Mariotti, A., 2006. Molecular
703 and isotopic study of lipids in particle size fractions of a sandy cultivated soil (Cestas cultivation
704 sequence, southwest France): Sources, degradation, and comparison with Cestas forest soil.
705 Organic Geochemistry 37, 20-44.

706 Reddé, M., Nuber, H.U., Jacomet, S., Schibler, J., Schucany, C., Schwarz, P.-A., Seitz, G.,
707 Ginella, F., Joly, M., Plouin, S., Hüster Plogmann, H., Petit, C., Popovitch, L., Schlumbaum,
708 A., Vandorpe, P., Viroulet, B., Wick, L., Wolf, J.-J., Gissinger, B., Ollive, V., Pellissier, J.,
709 2005. Oedenburg. Une agglomération d'époque romaine sur le Rhin supérieur: fouilles
710 françaises, allemandes et suisses à Biesheim-Kunheim (Haut-Rhin). Gallia 62, 215-277.

711 Reimer, P.J., Bard, E., Bayliss, A., Beck, J.W., Blackwell, P.G., Ramsey, C.B., Buck, C.E.,
712 Cheng, H., Edwards, R.L., Friedrich, M., Grootes, P.M., Guilderson, T.P., Haflidason, H.,
713 Hajdas, I., Hatté, C., Heaton, T.J., Hoffmann, D.L., Hogg, A.G., Hughen, K.A., Kaiser, K.F.,
714 Kromer, B., Manning, S.W., Niu, M., Reimer, R.W., Richards, D.A., Scott, E.M., Southon, J.R.,
715 Staff, R.A., Turney, C.S.M., van der Plicht, J., 2013. IntCal13 and Marine13 radiocarbon age
716 calibration curves 0-50,000 years cal BP. Radiocarbon 55, 1869-1887.

717 Rethemeyer, J., Kramer, C., Gleixner, G., Wiesenberg, G.L.B., Schwark, L., Andersen, N.,
718 Nadeau, M.-J., Grootes, P.M., 2004. Complexity of soil organic matter: AMS ¹⁴C analysis of
719 soil lipid fractions and individual compounds. Radiocarbon 46, 465-473.

720 Rethemeyer, J., Kramer, C., Gleixner, G., John, B., Yamashita, T., Flessa, H., Andersen, N.,
721 Nadeau, M.-J., Grootes, P.M., 2005. Transformation of organic matter in agricultural soils:
722 radiocarbon concentration versus soil depth. *Geoderma* 128, 94-105.

723 Rieley, G., 1994. Derivatization of organic compounds prior to gas chromatographic-
724 combustion-isotope ratio mass spectrometric analysis: identification of isotope fractionation
725 processes. *Analyst* 119, 915-919.

726 Ross, A.B., Shepherd, M.J., Schüpphaus, M., Sinclair, V., Alfaro, B., Kamal-Eldin, A., Åman,
727 P., 2003. Alkylresorcinols in cereal and cereal products. *Journal of Agricultural and Food*
728 *Chemistry* 51, 4111-4118.

729 Ross, A.B., Åman, P., Andersson, R., Kamal-Eldin, A., 2004. Chromatographic analysis of
730 alkylresorcinols and their metabolites. *Journal of Chromatography A* 1054, 157-164.

731 Ruas, M.-P., Marinval, P., 1991. Alimentation végétale et agriculture d'après les semences
732 archéologiques (de 9000 av. J.-C. au XV^e siècle). In: Guilaine, J. (Ed), *Pour une Archéologie*
733 *Agraire*. Armand Colin, pp. 409-439.

734 Ruff, M., Wacker, L., Gäggeler, H.W., Suter, M., Synal, H.-A., Szidat, S., 2007. A gas ion
735 source for radiocarbon measurements at 200 kV. *Radiocarbon* 49, 307-314.

736 Ruff, M., Fahrni, S., Gäggeler, H.W., Hajdas, I., Suter, M., Synal, H.-A., Szidat, S., Wacker,
737 L., 2010. On-line radiocarbon measurements of small samples using elemental analyzer and
738 MICADAS gas ion source. *Radiocarbon* 52, 1645-1656.

739 Schmidt, M.W.I., Torn, M.S., Abiven, S., Dittmar, T., Guggenberger, G., Janssens, I.A., Kleber,
740 M., Kögel-Knabner, I., Lehmann, J., Manning, D.A.C., Nannipieri, P., Rasse, D.P., Weiner, S.,
741 Trumbore, S.E., 2011. Persistence of soil organic matter as an ecosystem property. *Nature* 478,
742 49-56.

743 Schnell, G., Schaeffer, P., Tardivon, H., Motsch, E., Connan, J., Ertlen, D., Schwartz, D.,
744 Schneider, N., Adam, P., 2014. Contrasting diagenetic pathways of higher plant triterpenoids
745 in buried wood as a function of tree species. *Organic Geochemistry* 66, 107-124.

746 Schwartz, D., 1991. Intérêt de la mesure du $\delta^{13}\text{C}$ des sols en milieu naturel équatorial pour la
747 connaissance des aspects pédologiques et écologiques des relations savane-forêt. *Cahiers*
748 *ORSTOM, Série Pédologie* 26, 327-341.

749 Schwartz, D., 2012. Les temps du sol : interprétations temporelles de l'archivage pédologique
750 dans les approches paléoenvironmentalistes et géoarchéologiques. *Étude & Gestion des Sols*
751 19, 51-65.

752 Seitz, L.M., 1992. Identification of 5-(2-oxoalkyl)resorcinols and 5-(2-oxoalkenyl)resorcinols
753 in wheat and rye grains. *Journal of Agricultural and Food Chemistry* 40, 1541-1546.

754 Sistiaga, A., Mallol C., Galván, B., Summons, R.E., 2014. The Neanderthal meal: a new
755 perspective using faecal biomarkers. *PLOS ONE* 9, e101045.

756 Stuiver, M., Polach, H.A., 1977. Discussion: reporting of ^{14}C data. *Radiocarbon* 19, 355–363.

757 Synal, H.-A., Stocker, M., Suter, M., 2007. MICADAS: a new compact radiocarbon AMS
758 system. *Nuclear Instruments and Methods in Physics Research B* 259, 7-13.

759 Trendel, J.M., Schaeffer, P., Adam, P., Ertlen, D., Schwartz, D., 2010. Molecular
760 characterisation of soil surface horizons with different vegetation in the Vosges Massif (France).
761 *Organic Geochemistry* 41, 1036-1039.

762 Tulloch, A.P., 1982. Epicuticular waxes of *Panicum miliaceum*, *Panicum texanum* and *Setaria*
763 *italica*. *Phytochemistry* 21, 2251-2255.

764 van Bergen, P.F., Bull, I.D., Poulton, P.R., Evershed, R.P., 1997. Organic geochemical studies
765 of soils from the Rothamsted classical experiments – I. Total lipid extracts, solvent insoluble
766 residues and humic acids from Broadbalk Wilderness. *Organic Geochemistry* 26, 117-135.

767 van der Merwe, N.J., Vogel, J.C., 1983. Recent carbon isotope research and its implications for
768 African archaeology. *African Archaeological Review* 1, 35-56.

769 Vandorpe, P., Jacomet, S., 2011. Plant economy and environment. In: Vandorpe, P., Plant
770 macro remains from the 1st and 2nd Cent. A.D. in Roman Oedenburg/Biesheim-Kunheim (F).
771 Methodological aspects and insights into local nutrition, agricultural practices, import and the
772 natural environment. Habilitation Thesis, University of Basel, Switzerland.

773 Vandorpe, P., Wick, L., 2015. Food plants in anthropogenic sediments as represented in pollen
774 and macrofossil spectra: a case study on waterlogged Roman sediments from Switzerland and
775 Alsace, France. *Vegetation History and Archaeobotany* 24, 135-142.

776 Wacker, L., Němec, M., Bourquin, J., 2010a. A revolutionary graphitisation system: fully
777 automated, compact and simple. *Nuclear Instruments and Methods in Physics Research B* 268,
778 931-934.

779 Wacker, L., Bonani, G., Friedrich, M., Hajdas, I., Němec, M., Ruff, M., Suter, M., Synal, H.-
780 A., Vockenhuber, C., 2010b. MICADAS: routine and high-precision radiocarbon dating.
781 *Radiocarbon* 52, 252-262.

782 Wang, J., Liu, L., Ball, T., Yu, L., Li, Y., Xing, F., 2016. Revealing a 5,000-y-old beer recipe
783 in China. *Proceedings of the National Academy of Sciences USA* 113, 6444-6448.

784 Wiesenberg, G.L.B., Schwarzbauer, J., Schmidt, M.W.I., Schwark, L., 2004. Source and
785 turnover of organic matter in agricultural soils derived from *n*-alkane/*n*-carboxylic acid
786 compositions and C-isotope signatures. *Organic Geochemistry* 35, 1371-1393.

787 Wiethold, J., 2000. Macro-restes végétaux imbibés et carbonisés du premier âge du Fer
788 provenant de fouille sauvetage de chantier 'les résidences du vignoble' à Hattstatt-
789 'Ziegelscheuer' (Haut-Rhin). In: Dumont, A., Treffort, J.M. (Eds), Hattstatt « Ziegelsheuer »
790 « Les résidences du vignoble », 68 – Haut-Rhin, Puits à eau protohistoriques (Bronze Ancien
791 et Hattstatt C). Afan Grand-Est, Strasbourg.

792 Winata, A., Lorenz, K., 1997. Effects of fermentation and baking of whole wheat and whole
793 rye sourdough breads on cereal alkylresorcinols. *Cereal Chemistry* 74, 284-287.

794

795 **Figure captions:**

796 **Fig. 1.** Photos and cross-sections of selected archaeological structures unearthed at Obernai
797 (Alsace, NE France); a, Silo 1330; b, Gaulish pit; c, Roman Gaul latrines; AL, Archaeological
798 layer; LS, loess substrate. Photos and drawings by Clément Féliu (INRAP).

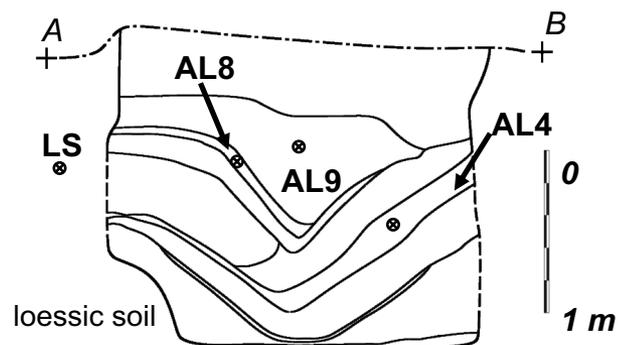
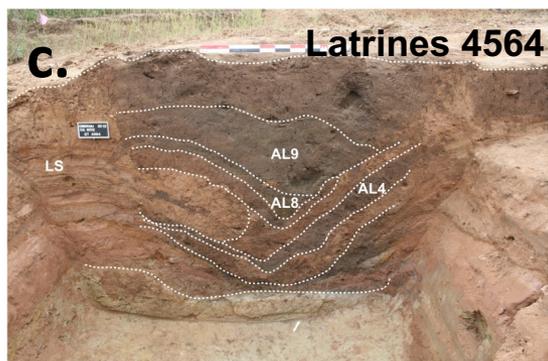
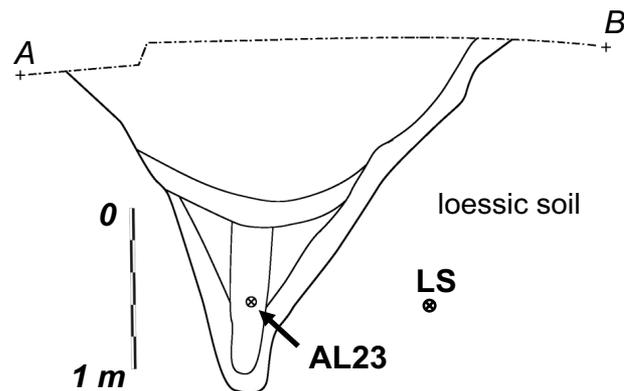
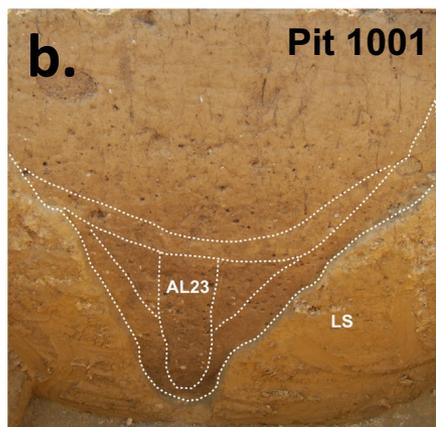
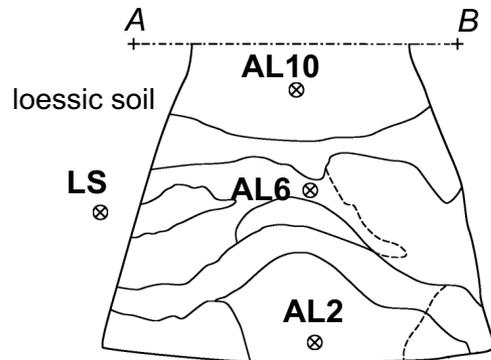
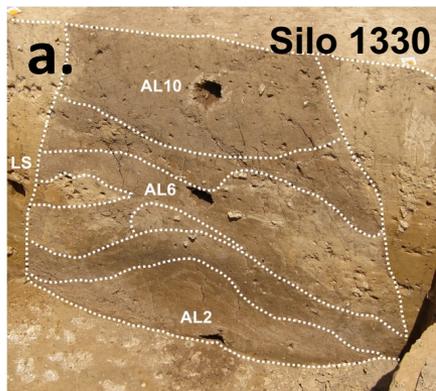
799 **Fig. 2.** Gas chromatograms (GC-MS, conditions B) of the apolar fraction from the lipid extract
800 from (a) silo 1330.AL2, (b) silo 1330.LS, (c) pit 1001.AL23 and (d) latrines 4564.AL8 (Obernai,
801 NE France). Carboxylic acids are analysed as methyl esters and alcohols as acetates. Numbers
802 refer to structures in Appendix A.

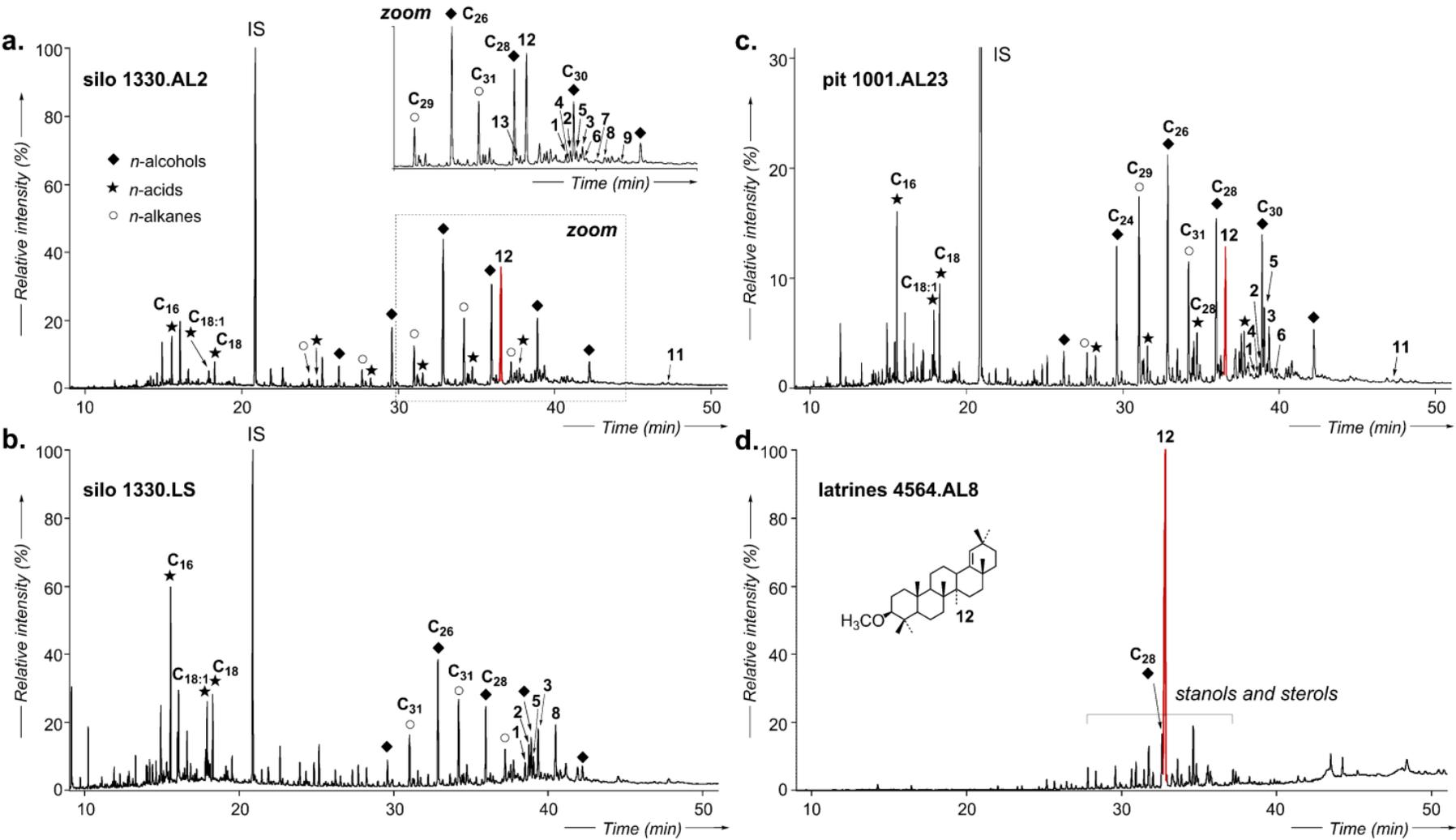
803 **Fig. 3.** Amount of miliacin ($\mu\text{g/g}$ soil) in the soils collected within the archaeological layers of
804 the silos (AL) and the Gaulish pit as well as in their respective loess substrates (LS).

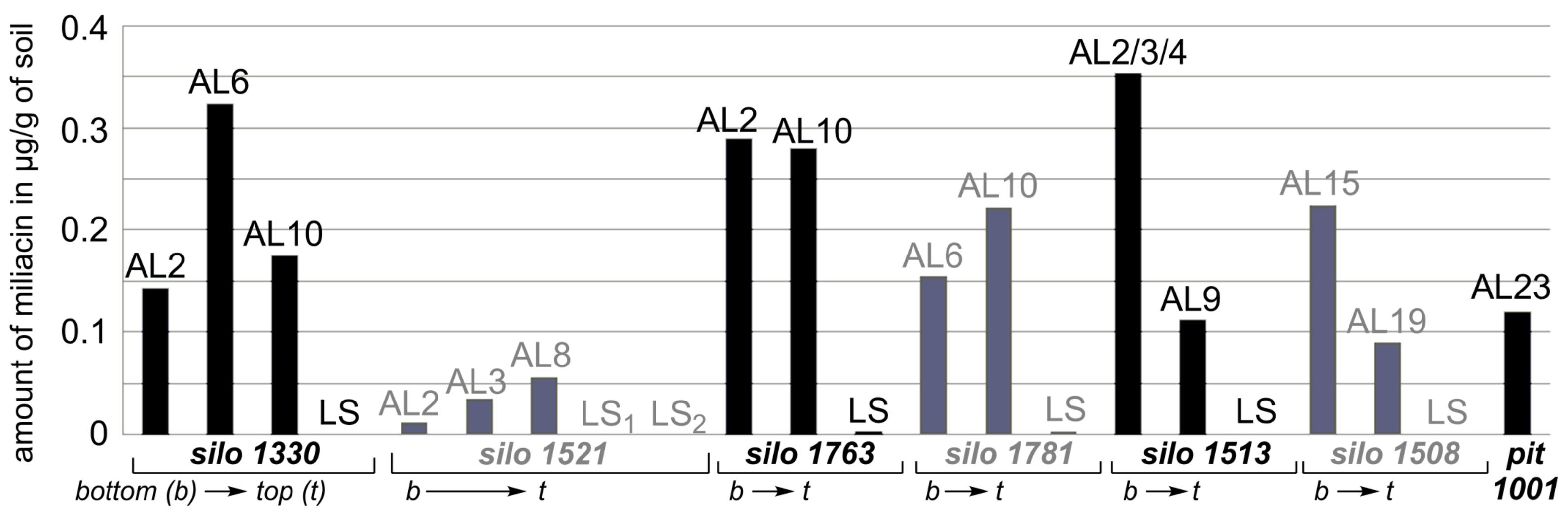
805 **Fig. 4.** Graphical representation of ^{14}C age cal. BP obtained for isolated miliacin samples (in
806 blue), lipid extracts (in red), soil organic matter (in grey), carbonized grains of millet (in green)
807 in respect to the age of the archaeological structures determined by ceramology. The non-
808 calibrated ^{14}C ages are marked with a black cross (for standard deviation, see Table 3).

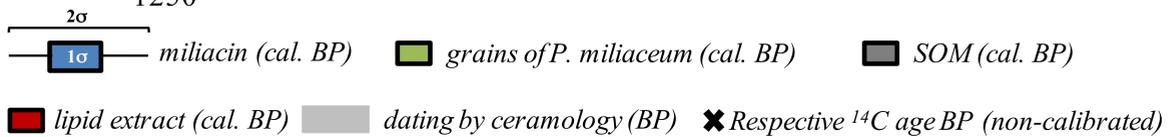
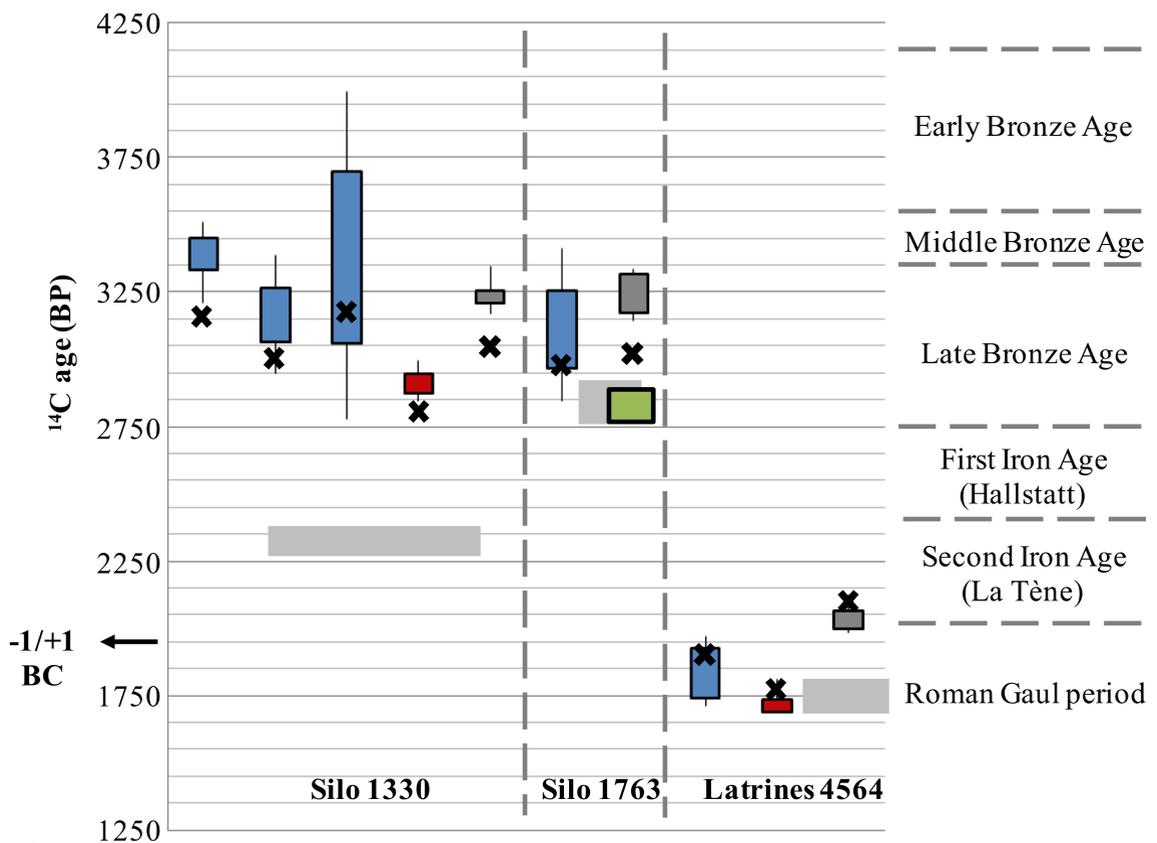
809

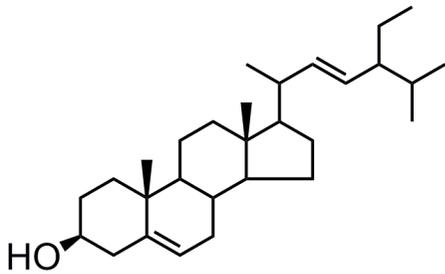
810



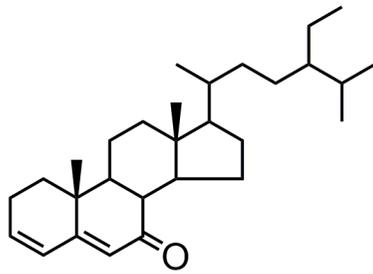




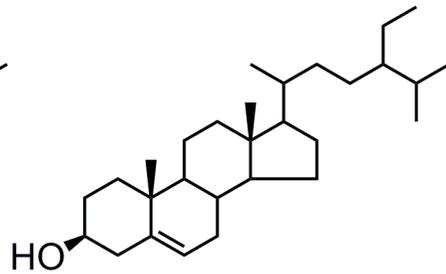




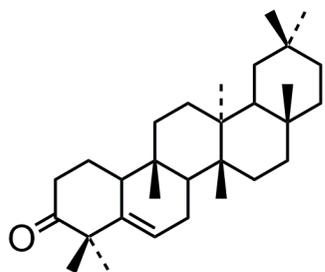
1



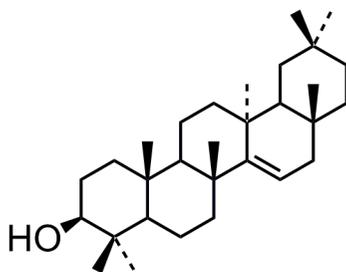
2



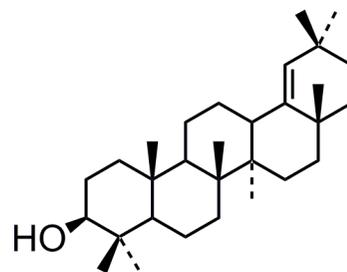
3



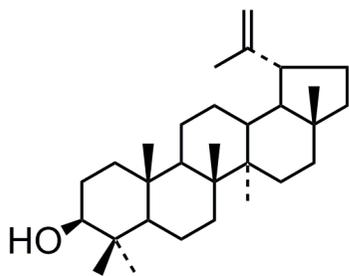
4



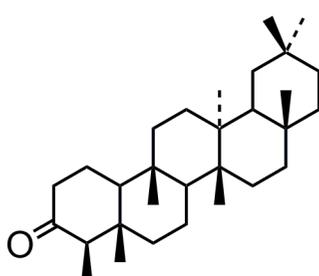
5



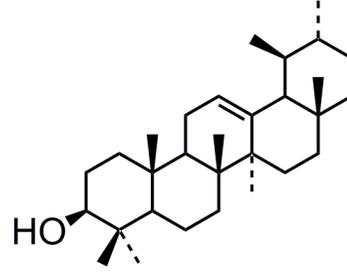
6



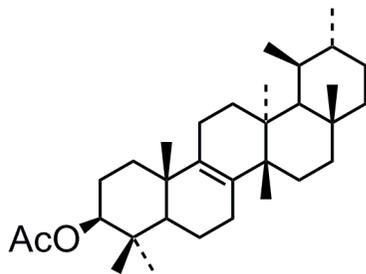
7



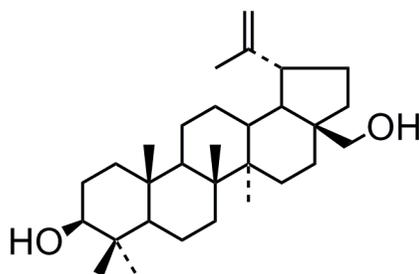
8



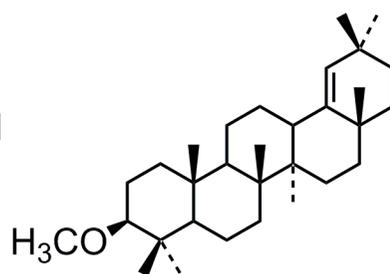
9



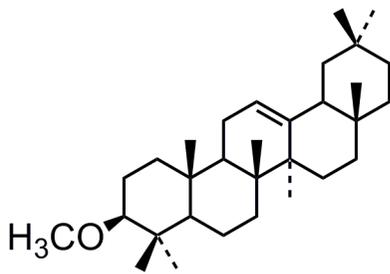
10



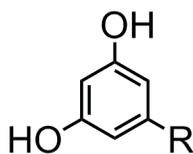
11



12



13



alkyresorcinols

R: Alkyl chains
(odd carbon numbered,
typically C₁₅-C₂₅)

Table 1

Archaeological structures sampled for lipid analysis and age of the structures determined from ceramology.

	Archaeological age	Historical period	Sampling level ^a
Silos			
1330	400-260 BC	La Tène B	AL2, AL6, AL10, LS
1508	400-260 BC	La Tène B	AL15, AL19, LS
1513	400-260 BC	La Tène B	AL2/3/4, AL9, LS
1521	150-30 BC	La Tène D	AL2, AL3, AL8, LS1, LS2
1763	950-800 BC	Late Bronze Age	AL2, AL10, LS
1781	400-260 BC	La Tène B	AL6, AL10, LS
1817	4600-4300 BC	Neolithic period	AL3, AL8, AL9, LS1, LS2
Latrines			
4564	End of 2 nd c. AD – beginning of the 3 rd c. AD	Roman period	AL4, AL8, AL9, LS
Gaulish pit			
1001	150-30 BC	La Tène D	AL23, LS

^a Location within the archaeological layers (AL) of the structures and in the loess substrates (LS) are illustrated in Fig. 1 and Fig. S1 (Supplementary data).

Table 2

Stable carbon isotopic composition ($\delta^{13}\text{C}$, ‰ vs. VPDB) and standard deviation ($\pm\sigma$) of individual lipids from selected silo samples (AL) and one loess substrate sample (LS ; n.d., not determined).

$\delta^{13}\text{C}$	1330.AL2	1508.AL15	1508.LS	1513.AL9	1521.AL8	1817.AL3	1763.AL2	$\delta^{13}\text{C}$	1508.AL15	1513.AL9	1521.AL8	1763.AL2
<i>n</i> -Alkanes								<i>n</i> -Acids ^a				
C ₂₅	-31.8 (\pm 0.7)	-32.1 (\pm 0.5)	-33.1 (\pm 0.4)	-32.5 (\pm 0.2)	-31.8 (\pm 0.4)	-31.8 (\pm 0.5)	-29.9 (\pm 0.5)	C ₂₀	-32.6 (\pm 0.4)	n.d.	n.d.	-31.6 (\pm 0.2)
C ₂₇	-31.5 (\pm 0.6)	-31.9 (\pm 0.2)	-32.1 (\pm 0.3)	-31.9 (\pm 0.2)	-31.7 (\pm 0.4)	-32.5 (\pm 0.1)	-30.4 (\pm 0.2)	C ₂₂	-33.8 (\pm 0.2)	n.d.	n.d.	-31.7 (\pm 0.2)
C ₂₉	-33.5 (\pm 0.5)	-34.0 (\pm 0.3)	-33.6 (\pm 0.1)	-33.9 (\pm 0.1)	-33.5 (\pm 0.3)	-33.5 (\pm 0.2)	-32.5 (\pm 0.2)	C ₂₄	-32.8 (\pm 0.2)	n.d.	n.d.	-32.8 (\pm 0.3)
C ₃₁	-34.7 (\pm 0.2)	-34.5 (\pm 0.2)	-34.2 (\pm 0.1)	-35.4 (\pm 0.5)	-34.1 (\pm 0.5)	-34.3 (\pm 0.2)	-33.4 (\pm 0.2)	C ₂₆	-33.7 (\pm 0.4)	n.d.	n.d.	-34.7 (\pm 0.4)
C ₃₃	-35.5 (\pm 0.3)	-33.0 (\pm 0.1)	-33.5 (\pm 0.3)	-33.9 (\pm 0.2)	-34.5 (\pm 1.0)	-34.5 (\pm 0.3)	-32.9 (\pm 0.4)	C ₂₈	-34.6 (\pm 0.2)	n.d.	n.d.	-36.3 (\pm 0.4)
<i>n</i> -Alcohols ^a								C ₃₀	n.d.	n.d.	n.d.	-36.2 (\pm 0.4)
C ₂₂	-33.6 (\pm 0.6)	-33.8 (\pm 0.4)	-33.6 (\pm 0.3)	-32.6 (\pm 0.2)	-33.6 (\pm 0.4)	-33.8 (\pm 0.4)	-33.4 (\pm 0.3)	C ₃₂	-34.9 (\pm 0.7)	n.d.	n.d.	-37.0 (\pm 0.5)
C ₂₄	-33.6 (\pm 0.3)	-33.9 (\pm 0.1)	-34.3 (\pm 0.2)	-33.7 (\pm 0.4)	-32.8 (\pm 0.1)	-33.4 (\pm 0.4)	-34.8 (\pm 0.2)	Miliacin (12)				
C ₂₆	-35.9 (\pm 0.2)	-36.2 (\pm 0.1)	-35.4 (\pm 0.1)	-36.2 (\pm 0.2)	-36.1 (\pm 0.2)	-34.9 (\pm 0.2)	-36.8 (\pm 0.2)		-21.6 (\pm 0.1)	-21.5 (\pm 0.1)	-21.3 (\pm 0.3)	n.d.
C ₂₈	-36.2 (\pm 0.2)	-36.1 (\pm 0.1)	-35.6 (\pm 0.1)	-36.1 (\pm 0.2)	-35.8 (\pm 0.2)	-35.3 (\pm 0.6)	-35.5 (\pm 0.1)					
C ₃₀	-34.6 (\pm 0.5)	-34.9 (\pm 0.2)	-35.6 (\pm 0.5)	-35.1 (\pm 0.3)	-35.8 (\pm 0.2)	-35.3 (\pm 0.3)	n.d.					
C ₃₂	-33.6 (\pm 0.7)	-31.4 (\pm 0.2)	-32.6 (\pm 0.4)	-33.1 (\pm 0.4)	-32.7 (\pm 0.1)	-35.4 (\pm 0.2)	-33.2 (\pm 0.4)					

^a Values corrected for derivatization agent [$\text{Si}(\text{CH}_3)_3$ and CH_3 for the *n*-alcohols and *n*-acids, respectively].

Table 3

¹⁴C age (in BP^a and cal. BC/AD^b) of soil organic matter (SOM), lipid extract (LE) and isolated miliacin (**12**) from selected soil samples.

Sample	Amount of soil	SOM (decarbonated soil) Age ¹⁴ C BP ^a (±1σ)	Cal. age ¹⁴ C (68%) ^b	Cal. age ¹⁴ C (95%) ^b
1330.AL6 ^c	230 mg	3042 (± 28)	1381-1343 BC ^c (27%) 1307-1260 BC (38%)	1396-1218 BC (95%)
1330.LS ^d	140 mg	7597 (± 54)	6492-6412 BC (68%)	6589-6378 BC (95%)
1763.AL2 ^c	200 mg	3022 (± 23)	1366-1221 BC (68%)	1386-1195 BC (95%)
4564.AL8 ^d	145 mg	2072 (± 27)	154-136 BC (11%) 114-46 BC (57%)	175-36 BC (92%)
Sample	Amount of LE	LE Age ¹⁴ C BP (±1σ)	Cal. age ¹⁴ C (68%)	Cal. age ¹⁴ C (95%)
1330.AL6	1.3 mg	2806 (± 28)	996-922 BC (68%)	1044-896 BC (95%)
1330.LS ^e	0.8 mg	122 (± 27)	1685-1707 AD ^f (12%) 1833-1885 AD (31%)	1679-1764 AD (33%) 1801-1897 AD (48%) 1902-1940 AD (15%)
4564.AL8	1.5 mg	1784 (± 25)	215-260 AD (38%) 280-325 AD (30%)	138-264 AD (61%) 274-330 AD (34%)
Sample	Amount of 12	Miliacin (12) Age ¹⁴ C BP (±1σ)	Cal. age ¹⁴ C (68%)	Cal. age ¹⁴ C (95%)
1330.AL2	40 μg	3151 (± 66)	1501-1381 BC (55%)	1560-1259 BC (93%)
1330.AL6	27 μg	2999 (± 87)	1317-1116 BC (57%)	1438-996 BC (95%)
1330.AL10	38 μg	3175 (± 255)	1750-1110 BC (68%)	2044-826 BC (95%)
1763.AL2	15 μg	2967 (± 127)	1307-1015 BC (61%)	1461-894 BC (93%)
4564.AL8	300 μg	1906 (±50)	26-209 AD (68%)	19 BC-236 AD (95%)

^aBP: "Before Present" where the year 1950, when nuclear weapons testing changed the ¹⁴C concentration in the atmosphere, is the reference (0 BP); ^bthe radiocarbon content within the atmosphere having varied over time, the calibration atmospheric curve used for the study includes important fluctuations (IntCal13; Reimer et al., 2013). Hence, the ¹⁴C age expressed in BP, i.e. before 1950 AD) can be associated with several possible ranges of time for the samples (cal. ¹⁴C age BC). The results with the highest statistical probability are reported here and used in Fig. 4, whereas the complete ¹⁴C dating data are reported in Table S3 (Supplementary material); ^cmeasure performed on soil sample after solvent extraction and decarbonation; ^dmeasure achieved on decarbonated soil sample; ^eBC: Before Christ; ^fA.: Anno Domini; ^gsample likely contaminated by input from recent organic material.

Highlights

- Miliacin (millet biomarker) found in paleosoils filling archaeological structures. •It was ^{14}C -dated after chromatographic isolation. • Hence, millet cropping was dated back to the Bronze Age. •Hollow structures act as “pedological traps” by sealing ancient cultivated soils. •The $\delta^{13}\text{C}$ lipid signature was used to decipher the C_3 vs. C_4 plant input to SOM.

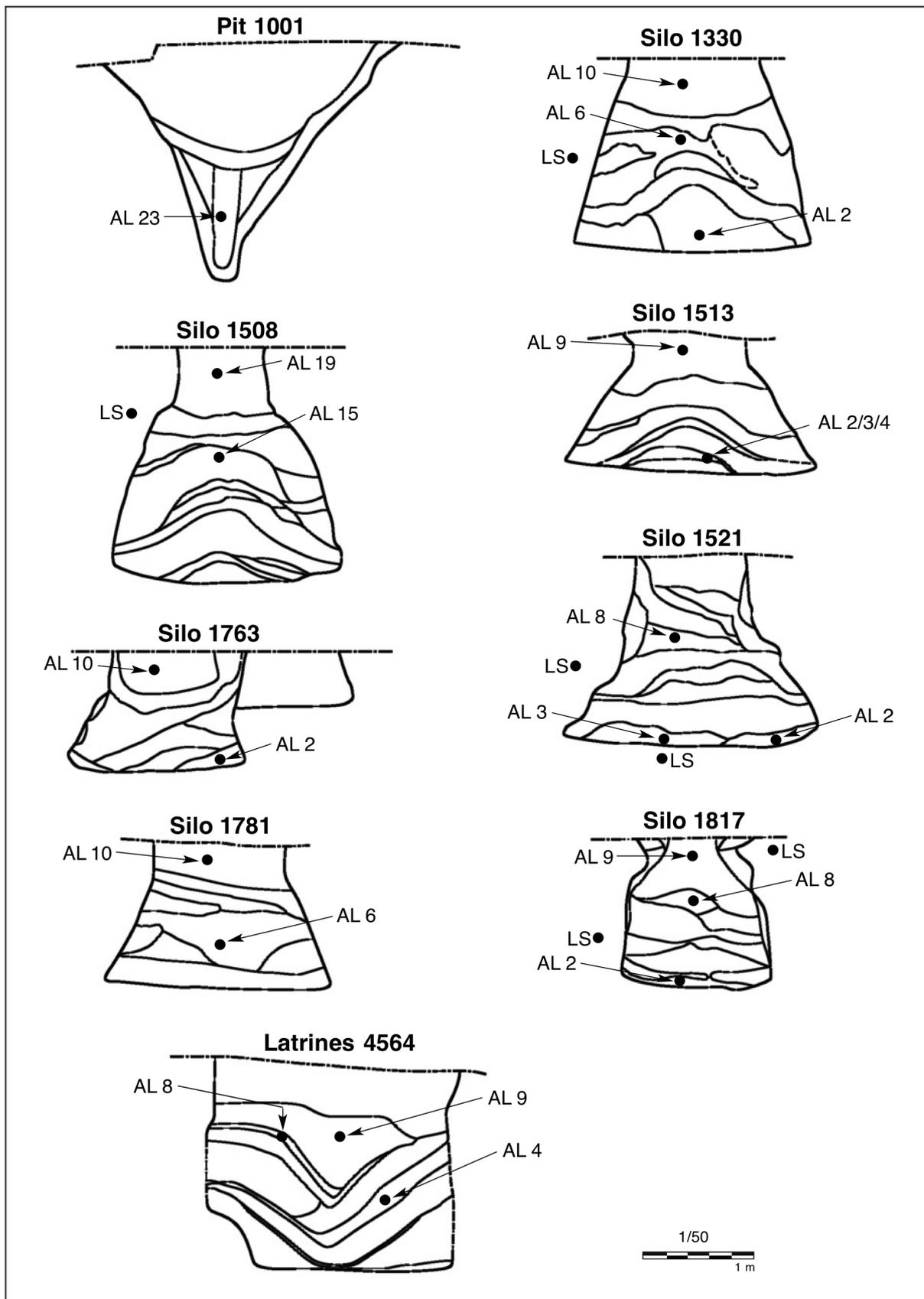


Fig. S1. Archaeological structures and location of samples in the archaeological layers (AL) and loess substrates (LS). Drawing modified after Clément Féliu (INRAP).

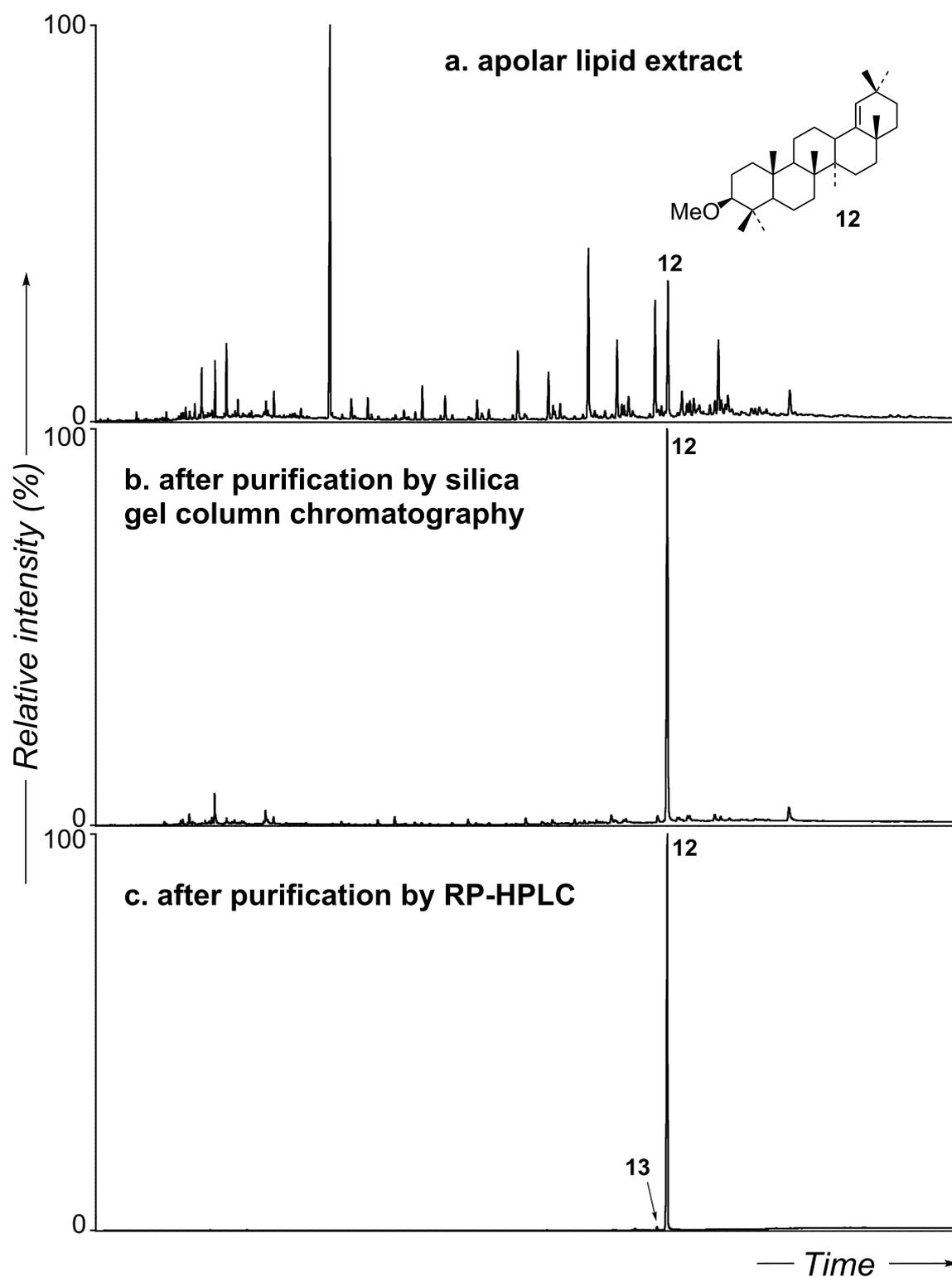


Fig. S2. Gas chromatograms (GC-MS) showing (a) apolar lipid extract from silo 1330.AL2, (b) miliacin-enriched fraction after a separation step using silica gel column chromatography (fraction eluted with CH_2Cl_2 ; cf. Section 2.6) and (c) isolated miliacin (**12**) after last purification step with RP-HPLC.

Table S1

MS data (EI, 70 eV) of sterols and triterpenoids in lipid extracts from the archaeological soil samples.

Compound	MW ^a	Main MS fragments (<i>m/z</i>)	Reference(s)
Stigmasterol (Ac) 1	454	147 (52), 213 (18), 255 (48), 267 (17), 282 (16), 296 (16), 351 (10), 379 (12), 394 (100)	Isobe et al. (2002)
Stigmastadienone 2	410	174 (100), 269 (13), 395 (13), 410 (37)	
β -Sitosterol (Ac) 3	456	145 (38), 147 (59), 213 (20), 255 (30), 275 (24), 288 (29), 381 (28), 396 (100)	Isobe et al. (2002)
Glutinone 4	424	150 (22), 189 (37), 205 (30), 245 (10), 259 (100) , 274 (57)	Shiojima et al. (1992)
Taraxerol (Ac) 5	468	189 (42), 204 (100) , 218 (25), 257 (17), 269 (27), 329 (22), 344 (31), 453 (7), 468 (10)	Shiojima et al. (1992); Lavrieux et al. (2011)
Germanicol (Ac) 6	468	161 (15), 177 (81), 189 (100) , 204 (90), 205 (45), 218 (31), 231 (13), 393 (3), 408 (4), 453 (15), 468 (18)	Shiojima et al. (1992); Oyo-Ita et al. (2010)
Lupeol (Ac) 7	468	175 (38), 189 (100) , 203 (37), 204 (40), 218 (28), 249 (17), 453 (6), 468 (19)	Shiojima et al. (1992); Stiti et al. (2007)
Friedelin 8	426	109 (89), 123 (100) , 125 (52), 163 (56), 191 (48), 205 (45), 246 (44), 273 (39), 302 (17), 341 (8), 411 (17), 426 (23)	Budzikiewicz et al. (1963)
α -Amyrin (Ac) 9	468	189 (41), 203 (28), 218 (100) , 249 (5), 408 (3), 468 (4)	Shiojima et al. (1992); Lavrieux et al. (2011)
Baurenyl acetate 10	468	229 (100) , 241 (20), 289 (94), 301 (12), 393 (7), 408 (7), 453 (8), 468 (5)	Lavrieux et al. (2011)
Betulin (di-Ac) 11	526	175 (24), 187 (64), 189 (100) , 191 (59), 203 (41), 216 (28), 249 (6), 393 (7), 423 (19), 451 (9), 466 (33), 526 (1)	Stiti et al. (2007)
Miliacin 12	440	175 (32), 177 (64), 189 (100) , 204 (80), 218 (36), 231 (22), 393 (3), 425 (18), 440 (18)	Jacob et al. (2005)
Isosawamilletin 13	440	175 (7), 189 (14), 203 (50), 218 (100) , 440 (1)	Jacob et al. (2005)

^a Alcohols analyzed as acetate derivatives (Ac).

Table S2

¹⁴C age (BP^a and cal. BC/AD^b) and fraction modern (F¹⁴C)^c determined for soil organic matter (SOM), lipid extracts (LE) and isolated miliacin (**12**) from selected silo samples (AL) and loess substrate samples (LS).

Sample	Number (ETH)	Amount of soil	SOM (decarbonated soil)			
			Age ¹⁴ C BP ^a (± 1σ)	F ¹⁴ C ^e (± 1σ) ^c	Cal. age ¹⁴ C (68.2%) ^b	Cal. age ¹⁴ C (95.4%) ^b
1330.AL6 ^d	ETH-65607	230 mg	3042 (± 28)	0.762 (± 0.003)	1381-1343 BC ^f (27.0%) 1307-1260 BC (37.6%) 1242-1235 BC (3.6%)	1396-1218 BC (95.4%)
1330.LS ^c	ETH-65606	140 mg	7597 (± 54)	0.388 (± 0.003)	6492-6412 BC (68.2%)	6589-6378 BC (95.4%)
1763.AL2 ^d	ETH-66449	200 mg	3022 (± 23)	0.687 (± 0.002)	1366-1221 BC (68.2%)	1386-1195 BC (95.4%)
4564.AL8 ^c	ETH-65605	145 mg	2072 (± 27)	0.773 (± 0.003)	154-136 BC (11.5%) 114-46 BC (56.7%)	175-36 BC (91.8%) 31-20 BC (1.7%) 12-1 BC (1.9%)
Sample	Number (ETH)	Amount of LE	LE			
			Age ¹⁴ C BP (± 1σ)	F ¹⁴ C (± 1σ)	Cal. age ¹⁴ C (68.2%)	Cal. age ¹⁴ C (95.4%)
1330.AL6	ETH-59648	1.3 mg	2806 (± 28)	0.705 (± 0.002)	996-922 BC (68.2%)	1044-896 BC (95.4%)
1330.LS ^h	ETH-63066	0.8 mg	122 (± 27)	0.985 (± 0.003)	1685-1707 AD ^g (12.3%) 1719-1732 AD (7.0%) 1808-1826 AD (9.4%) 1833-1885 AD (31.5%) 1913-1927 AD (8.0%)	1679-1764 AD (32.7%) 1801-1897 AD (47.8%) 1902-1940 AD (14.9%)
4564.AL8	ETH-63064	1.5 mg	1784 (± 25)	0.801 (± 0.003)	215-260 AD (37.9%) 280-325 AD (30.3%)	138-264 AD (61.0%) 274-330 AD (34.4%)
Sample	Number (ETH)	Amount of 12	Miliacin (12)			
			Age ¹⁴ C BP (± 1σ)	F ¹⁴ C (± 1σ)	Cal. age ¹⁴ C (68.2%)	Cal. age ¹⁴ C (95.4%)
1330.AL2	ETH-59645	40 μg	3151 (± 66)	0.676 (± 0.006)	1501-1381 BC (55.4%) 1342-1306 BC (12.8%)	1607-1582 BC (2.1%) 1560-1259 BC (92.8%) 1242-1234 BC (0.5%)
1330.AL6	ETH-59646	27 μg	2999 (± 87)	0.688 (± 0.008)	1386-1340 BC (11.5%) 1317-1116 BC (56.7%)	1438-996 BC (95.4%)
1330.AL10	ETH-59647	38 μg	3175 (± 255)	0.674 (± 0.021)	1750-1110 BC (68.2%)	2124-2091 BC (0.7%) 2044-826 BC (94.7%)
1763.AL2	ETH-63065	15 μg	2967 (± 127)	0.691 (± 0.011)	1381-1342 BC (6.9%) 1307-1015 BC (61.3%)	1496-1474 BC (1.0%) 1461-894 BC (93.5%) 872-852 BC (0.8%)
4564.AL8	ETH-65602	300 μg	1906 (± 50)	0.789 (± 0.005)	26-209 AD (68.2%)	19 BC-236 AD (95.4%)

^aBP: "Before Present" where the year 1950, when nuclear weapons testing changed the ¹⁴C concentration in the atmosphere, is the reference (0 BP); ^bthe radiocarbon content within the atmosphere having varied over time, the calibration atmospheric curve used includes important fluctuations (IntCal13; Reimer et al., 2013). Hence, the ¹⁴C age expressed in BP (i.e. before 1950 AD) can be associated with several possible ranges of time for the samples (cal. ¹⁴C age BC); ^cF¹⁴C: fraction modern, F¹⁴C = exp(-¹⁴Cage/8033); see Reimer et al. (2004); ^dmeasure performed on soil sample after extraction and decarbonation; ^emeasure achieved on decarbonated soil sample; ^fBC: before Christ; ^gAD: Anno Domini; ^hsample likely contaminated by input from recent organic material.

References

- Budzikiewicz, H., Wilson, J.M., Djerassi, C., 1963. Mass spectrometry in structural and stereochemical problems. XXXII. Pentacyclic triterpenes. *Journal of the American Chemical Society* 85, 3688-3699.
- Isobe, K.O., Tarao, M., Zakaria, M.P., Chiem, N.H., Minh, L.Y., Takada, H., 2002. Quantitative application of fecal sterols using gas chromatography-mass spectrometry to investigate fecal pollution in tropical waters: Western Malaysia and Mekong Delta, Vietnam. *Environmental Science & Technology* 36, 4497-4507.
- Jacob, J., Disnar, J.-R., Boussafir, M., Albuquerque, A.L.S., Sifeddine, A., Turcq, B., 2005. Pentacyclic triterpene methyl ethers in recent lacustrine sediments (Lagoa do Caçó, Brazil). *Organic Geochemistry* 36, 449-461.
- Lavrieux, M., Jacob, J., Le Milbeau, C., Zocatelli, R., Masuda, K., Bréheret, J.-G., Disnar, J.-R., 2011. Occurrence of triterpenyl acetates in soil and their potential as chemotaxonomical markers of Asteraceae. *Organic Geochemistry* 42, 1315-1323.
- Oyo-Ita, O.E., Ekpo, B.O., Oros, D.R., Simoneit, B.R.T., 2010. Occurrence and sources of triterpenoid methyl ethers and acetates in sediments of the cross-river system, Southeast Nigeria. *International Journal of Analytical Chemistry*, doi:10.1155/2010/502076.
- Reimer, P.J., Brown, T.A., Reimer, R.W., 2004. Discussion: reporting and calibration of post-bomb ^{14}C data. *Radiocarbon* 46, 1299-1304.
- Shiojima, K., Arai, Y., Masuda, K., Takase, Y., Ageta, T., Ageta, H., 1992. Mass spectra of pentacyclic triterpenoids. *Chemical and Pharmaceutical Bulletin* 40, 1683-1690.
- Stiti, N., Triki, S., Hartmann, M.-A., 2007. Formation of triterpenoids throughout *Olea europaea* fruit ontogeny. *Lipids* 42, 55-67.